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Guanine Radicals Induced in DNA by Low-Energy Photoionization

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CONSPECTUS. Guanine (**G**) radicals are precursors to DNA oxidative damage, correlated with carcinogenesis and aging. During the past few years, we demonstrated clearly an intriguing effect: **G** radicals can be generated upon direct absorption of UV radiation with energy significantly lower than the **G** ionization potential. Using nanosecond transient absorption spectroscopy, we studied the primary species, ejected electrons and guanine radicals, which result from photoionization of various DNA systems in aqueous solution.

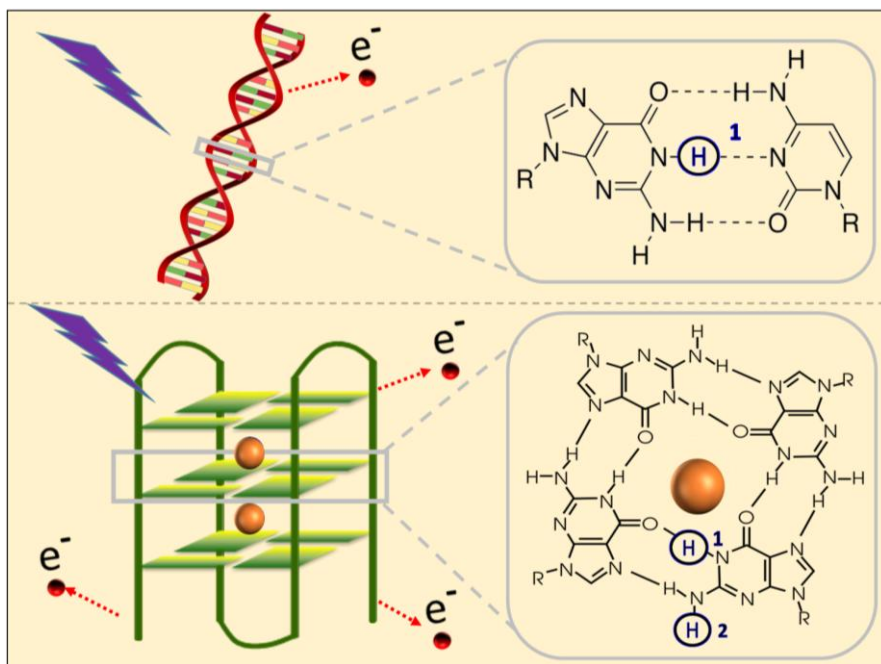
The DNA propensity to undergo electron detachment at low photon energies greatly depends on its secondary structure. Non-detected for monomers or unstacked oligomers, it may be one order of magnitude higher for **G**-quadruplexes than for duplexes. The experimental results point toward non-vertical processes, associated with the relaxation of electronic excited states. Theoretical studies are required to validate the mechanism and determine the factors that enter in play. In any case, such a mechanism, which may be operative over a broad excitation wavelength range, explains the occurrence of oxidative damage observed upon UVB and UVA irradiation.

Quantification of **G** radical populations and their time evolution questioned some widespread views. It appeared that **G** radicals may be generated with the same probability as pyrimidine dimers, which are considered to be the major lesions induced upon absorption of low-energy UV radiation by DNA. As the most important part of the initially created radical cations undergoes deprotonation, the vast majority of the final reaction products is expected to stem from long-lived deprotonated radicals. Consequently, the widely used oxidation marker 8-oxodG is not representative of the extent of oxidative damage when this is triggered by generation of radical cations.

Beyond the biological consequences, photo-generation of electron holes in **G**-quadruplexes may inspire applications in nano-electronics; although four-stranded structures are currently studied as molecular wires, their behavior as photoconductors has not been explored so far.

In the present Account, after highlighting some key experimental issues, we first describe the photo-ionization process and, then, we focus on radicals. We use as show-cases new results obtained for genomic DNA and *Oxytricha* **G**-quadruplexes. Generation and reaction dynamics of **G** radicals in these systems provide a representative picture of the phenomena reported previously for duplexes and **G**-quadruplexes, respectively.

CONSPECTUS Figure



1. Introduction

UV radiation instigates oxidative damage to DNA in an indirect or a direct way.¹ The former mechanism involves mediation of other molecules that absorb light and, subsequently, oxidize DNA through release of reactive oxygen species or photosensitized electron abstraction. In the latter, a photon directly absorbed by DNA provokes photo-ionization: an electron is ejected and a hole, that is a radical cation, is created. Regardless the mechanism and the initial location of the electron hole, after charge transfer,²⁻⁸ it reaches ultimately a guanine site. This happens because the guanine has the lowest oxidation potential compared to the other nucleobases, sugar and phosphate moieties composing DNA.⁹ The present Account describes guanine radical generation via low-energy photo-ionization and discusses their evolution.

As the vertical ionization potential (VIP) of the various DNA components is higher than 6.6 eV^{10,11} and excitation at the lowest absorption band (Figure 1a) is not expected to provoke one-photon ionization. Therefore, this phenomenon was studied by exciting at 193 nm (6.4 eV).¹²⁻¹⁴ Yet, more than a decade ago, two studies reported electron ejection from DNA oligomers, both in aqueous solution¹⁵ and in the gas phase,¹⁶ at much lower energy (266 nm, 4.7 eV); but these results appeared rather as a curiosity, connected with laser experiments. More recently, some of us showed that low-intensity continuous irradiation of purified genomic DNA at 554 and 295 nm gives rise not only to the well-known oxidation marker 8-oxo-7,8-dihydro-2'-deoxyguanosine (8-oxodG), but also to polyamine-guanine adducts.¹⁷ While 8-oxodG can be produced by several pathways, including DNA-mediated singlet oxygen generation,^{18,19} the detected adducts are typical of a mechanism involving the guanine radical cation (G^+)[•]. In parallel, we have undertaken a study aiming at the quantitative characterization of the related primary species, ejected electrons and guanine radicals, by transient absorption spectroscopy using as excitation

source 5 ns laser pulses at 266 nm. Although nanosecond flash photolysis has been a laboratory tool for over half a century now, specific protocols had to be developed for these studies. The reason is that they require detection of very weak signals arising from DNA systems undergoing damage during the measurement.

Our photo-ionization experiments have also proven a unique tool to study the “intrinsic” reaction dynamics of radicals; the term “intrinsic” refers to the absence of any external additive other than the ingredients of the phosphate buffer used as a solvent, which do not react with **G** radicals. As guanine radical cations ($\text{G}^+\cdot$) are known to be metastable undergoing either hydration or loss of a proton,²⁰ the time-resolved spectra allow us to follow their transformation and evaluate the evolution of their populations. This is an important issue because deprotonated radicals give rise to lesions different from those stemming directly from ($\text{G}^+\cdot$).¹ Consequently, the final type of oxidative damage depends, among others, on the population of ($\text{G}^+\cdot$) that undergoes deprotonation.

Our studies revealed that the secondary DNA structure plays a key role both in the generation of guanine radicals by low-energy photo-ionization and in their evolution. Here, we discuss our previous studies on model duplexes²¹⁻²⁴ and present new results on calf thymus (**CT-DNA**), showing that genomic DNA exhibits the same behavior as small size systems. In addition, we exemplify the behavior of **G**-quadruplexes (**G4**), whose building blocks are guanine tetrads (Conspectus Figure), by presenting new data obtained for Oxytricha **G4**. The latter, obtained by association of two single strands GGGGTTTTGGGG (**OXY**),²⁵ confirm previous studies reported for monomolecular^{24,26,27} and tetramolecular systems.²⁸⁻³⁰ Our **G4** are formed and studied in the presence of Na^+ or K^+ cations. These cations, are not only part

of the bulk solvent; they are also located in the central cavity of **G4**, being constitutive elements of their structure.³¹ Therefore, when referring to **G4**, we always indicate the type of metal cations, for example: **OXY/Na⁺** and **OXY/K⁺**.

Key experimental points

Experimental details are given in the Supporting Information (SI-Section 1). Here, we simply highlight a few points that are crucial for these studies.

Materials. Both synthetic oligomers and genomic DNA were extensively purified. Moreover, the purity of the chemicals used for the preparation of the buffer was 99.99%. This is an important requirement for avoiding photo-ionization of impurities, on the one hand, and their reaction with radicals, on the other. The buffer (pH 7) was composed of an equimolar mixture of MH_2PO_4 , and M_2HPO_4 , where M^+ is either Na^+ or K^+ . In most experiments, their concentration was high ($0.15 \text{ mol}\cdot\text{L}^{-1}$) so that to scavenge the ejected electrons,³² preventing their interaction with DNA. Furthermore, the corresponding high ionic strength helped stabilizing short duplexes. During the measurement, the solution was frequently renewed so that to avoid that successive laser pulses excite already damaged DNA; typically, for a complete study of a given system, 300-500 mL were needed.

Spectroscopic measurements. The challenge was to work with low-excitation intensities, leading to very weak transient absorption signals, so that to avoid two-photon ionization of the buffer, saturation effects in electron ejection (SI-Section 2), interaction among radicals and minimize two-photon ionization of DNA. Transient absorption signals were optimized by using orthogonal exciting and probing beams whose path-lengths though the sample were, respectively, 0.1 cm and 1 cm; the absorbance of the solution on the excitation side was 0.25 ± 0.02 , corresponding to a base concentration of the order of $10^{-4} \text{ mol}\cdot\text{L}^{-1}$. The incident pulse

intensity did not exceed, depending on the system, 2×10^6 or 3×10^6 $\text{W} \cdot \text{cm}^{-2}$ (SI-Section 2). In this way, the transient absorption decays are not influenced by the incident excitation intensity (Figure S2). Finally, the concentration of photons absorbed in the probed volume of the studied solution $[\text{h}\nu]$ was determined using a NIST traceable power-meter and counterchecked via an actinometer (SI-Section 1). In all experiments, the concentration of generated radicals was three orders of magnitude lower than the base concentration.

2. Characterization of ejected electrons

Electrons issued from photo-ionization in aqueous solution are known to be hydrated (e_{hyd}^-) and exhibit a broad absorption band peaking around 720 nm. Such a band is indeed present in the transient absorption spectrum recorded for **CT-DNA** at 30 ns, corresponding to the 30-ns time resolution of our experimental setup (Figure 1b). Based on this property and taking into account a maximum molar absorption coefficient ϵ of $19700 \text{ mol}^{-1} \cdot \text{L} \cdot \text{cm}^{-1}$ reported for e_{hyd}^- ,³³ we quantify the ejected electrons by probing their transient absorption on the sub-microsecond time-scale.

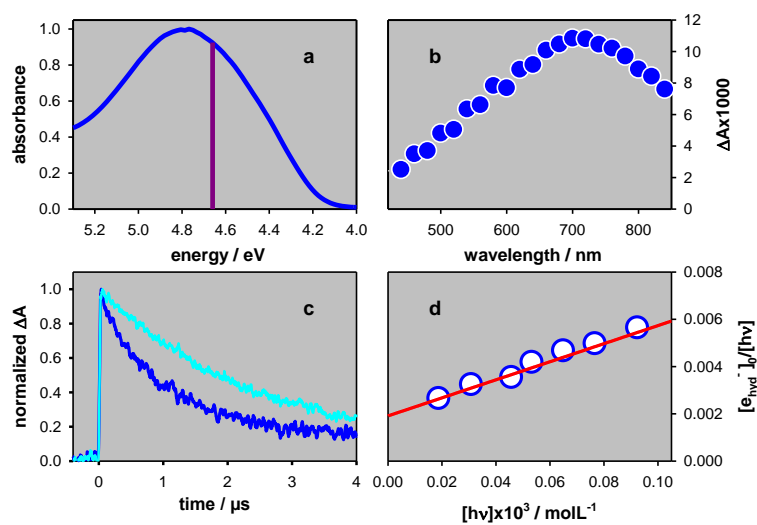


Figure 1. CT-DNA (blue) in phosphate buffer (0.015 mol·L⁻¹ NaH₂PO₄ / 0.015 mol·L⁻¹ Na₂HPO₄). (a) Steady-state absorption spectrum; the vertical violet line indicates the excitation wavelength at 266 nm. (b) Transient absorption spectrum recorded at 30 ns with incident excitation intensity of 2.0×10⁶ W·cm⁻². (c) Normalized transient absorption decay at 700 nm; it is compared with the corresponding signal obtained under the same conditions for a short duplex (cyan; **S1•S2**,²⁴ Table 1). (d) Ionization curve; [hv] and [e_{hyd}⁻]₀ denote, respectively, the concentration of absorbed photons per laser pulse and the initial concentration of hydrated electrons; the red line is the fit with a linear model function. (b-d): argon saturated solutions.

In our experimental conditions, e_{hyd}⁻ disappear within a few μs, reacting with H₂PO₄⁻ ions³² present in high concentration. Their decay rate is the same for all the small DNA systems (Figure S3), depending only on the buffer concentration. In contrast, for the same buffer concentration, e_{hyd}⁻ stemming from CT-DNA decay significantly faster (Figures 1c), revealing the existence of additional reaction paths, involving DNA.³⁴ Such reactions could be favored by the flexibility of the very long duplex,³⁵ increasing locally the encounter probability between nucleic acid components and e_{hyd}⁻ produced by the same helix.

Although we use low excitation intensities, two-photon ionization, resulting from absorption of a second photon by a long-lived electronic excited state, cannot be completely prevented. To disentangle one- and two-photon events, we vary the excitation intensity, thus changing the concentration of absorbed photons. Subsequently, we plot [e_{hyd}⁻]₀/[hv] versus [hv], obtaining the so-called ionization curve (Figure 1d). The determination of [e_{hyd}⁻]₀ is illustrated in Figure S3: the e_{hyd}⁻ decay is fitted with a model function A₁+A₀exp(-t/τ₁); subsequently, [e_{hyd}⁻]₀ is obtained from the A₀ value using the corresponding molar absorption coefficient. The fit of the experimental points in Figure 1d with a linear function [e_{hyd}⁻]₀/[hv] = φ₁ + α[hv] provides the quantum yield for one-photon ionization φ₁, while the contribution of two-photon ionization α[hv] is given by the slope of the curve.

We did not detect one-photon ionization (φ₁ ≤ 3×10⁻⁴) for mononucleotides and small single strands characterized by poor base stacking, such as the 6-mer telomere repeat TTAGGG²⁶ or

thymine single strands (dT)₂₀.¹⁵ In contrast, for adenine single strands, known to adopt a helical structure, a ϕ_1 value of $(1.0 \pm 0.1) \times 10^{-3}$ was found.²¹ Those determined for various duplexes and **G4** are gathered on Table 1. In the case of duplexes, ϕ_1 ranges from 1.0×10^{-3} to 2.0×10^{-3} .²¹⁻²⁴ This concerns also icosamers containing only adenines and thymines in alternating²² or homopolymeric sequence.²¹ Clearly higher values, which may approach 10^{-2} , are observed for **G4**.^{24,26,28-30} In this family, topology and molecularity seem to play a rather minor role: the behavior of tetramolecular **(TG₄T)₄/Na⁺** and monomolecular **TEL21/Na⁺** systems is similar although their guanines adopt, respectively, parallel and antiparallel orientations in respect to the glycosidic bonds.^{26,28} In contrast, the metal cation used in the buffer in which **G4** are formed and studied, has an important effect: the values found in the case of K⁺ are significantly higher compared to Na⁺.

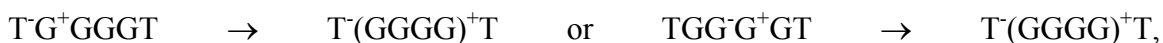
Table 1. Quantum yields ($\phi_1 \times 10^3$) of one-photon ionization at 266 nm and guanine/adenine radical half-lives (ms, in brackets) determined for duplexes and **G4** in phosphate buffer.

	M ⁺	CT-DNA	(GC) ₅ •(GC) ₅ ^a	S1•S2 ^b	(AT) ₁₀ •(AT) ₁₀ ^c	(A) ₂₀ •(T) ₂₀ ^d
duplexes	Na ⁺	2.0 ± 0.2 (3.7 ± 0.2)	1.2 ± 0.2 ²³ (2.5 ± 0.2)	2.1 ± 0.4 ²⁴ (4 ± 0.2)	1.5 ± 0.3 ²² (2.0 ± 0.1)	1.1 ± 0.3 ²¹ (4.0 ± 1.0)
	K ⁺	1.9 ± 0.1 (3.6 ± 0.2)	-	-	-	-
	M ⁺	TEL21 ^e	TEL25 ^f	OXY ^g	(TG ₄ T) ₄ ^h	
G4	Na ⁺	4.5 ± 0.6 ²⁶ (6.7 ± 0.5)	5.2 ± 0.3 ²⁴ (3.5 ± 0.2)	6.0 ± 0.2 (4.1 ± 0.2)	3.5 ± 0.5 ²⁸ (6.0 ± 0.5)	
	K ⁺	9.4 ± 0.1 ²⁷ (1.4 ± 0.1)	-	7.9 ± 0.1 (2.1 ± 0.2)	8.1 ± 0.5 ^{29,30} (3.1 ± 0.2)	

^{a)} alternating guanine-cytosine; ^{b)} S1: 5'-CGTACTCTTTGGTGGGTCGGTTCTTTCTAT-3' / S2: 3'-GCATGAGAAACCACCCAGCCAAGAAAGATA-5'; ^{c)} alternating adenine-thymine; ^{d)} homopolymeric adenine-thymine; ^{e)} monomolecular: GGG(TTAGGG)₃; ^{f)} monomolecular: TAGGG(TTAGGG)₃TT; ^{g)} bimolecular: 2×(GGGGTTTTGGGG); ^{h)} tetramolecular: 4×(TGGGGT).

3. Evidence for a non-vertical photo-ionization mechanism?

The effect of metal ions on the propensity of **G4** to undergo low-energy photo-ionization points towards non-vertical electron ejection. This conclusion was drawn, in particular, by comparing $(\text{TG}_4\text{T})_4/\text{Na}^+$ and $(\text{TG}_4\text{T})_4/\text{K}^+$,²⁸⁻³⁰ in which the static arrangement of nucleobases is the same. Although the VIP of these two systems, computed by quantum chemistry methods, are practically identical,¹⁹ we observe a twofold increase in ϕ_1 upon replacing Na^+ by K^+ cations. Hence, a complex mechanism, associated with the excited state relaxation, was proposed. The first step implies population of an excited charge transfer (CT) state, either between a thymine and a guanine or between two guanines. In a second step (charge separation), the positive charge is trapped by the guanine core, where it may be delocalized,^{11,36-38} while the negative one is born by a thymine. This process is tentatively described as:



where the radical sign was omitted for simplicity. Finally, electron ejection, assisted by conformational motions, occurs from the thymine anion, whose VIP in the gas phase is lower than 0.1 eV.³⁹ In the frame of such a scenario, Na^+ cations, smaller and more mobile than K^+ ,⁴⁰ allow the geometrical arrangement accompanying stabilization of CT states,⁴¹ and, hence, reducing the probability of charge separation.

Population of excited CT states both in duplexes⁴²⁻⁴⁴ and **G4**^{41,45,46} is well documented. The occurrence of charge separation along DNA strands has also been reported.⁴⁷ The higher ϕ_1 in **G4**, could be due to more efficient trapping of the positive charge within stacked guanine tetrads (Conspectus Figure); in this way, charge separation becomes more efficient, increasing the probability of electron ejection from a base located outside the guanine core. Another important difference is that guanine tetrads interact directly with dehydrated cations.³¹ Such interaction is

not encountered in duplexes, explaining why the ϕ_1 of **CT-DNA** is not significantly different in presence of Na^+ or K^+ cations in contrast to **G4** scaffolds (Table 1).

In contrast to the vertical ionization process, whose quantum yield decreases rapidly when moving away from the VIP, the proposed mechanism is expected to be operative over a broader range of excitation energy, provided that CT states are populated and charge separation achieved. This is supported by the fact that similar quantum yields for 8-oxodG formation were obtained for UVC and UVB irradiation.¹⁷ Furthermore, strand breaks were observed upon UVA irradiation,⁴⁸ where the weakly absorbing excited states^{49,50} have partial charge transfer character.^{50,51}

4. Radicals

General considerations. The spectral fingerprints of radicals are determined after the decay of e_{hyd}^- . We identify them by comparison with the corresponding spectra reported for monomeric guanosine derivatives (Figure 2a). We checked experimentally the validity of this comparison for $(\text{G}^+)^\bullet$, for which stronger interactions with neighboring bases are expected than for deprotonated radicals, because of the presence of a positive charge. To this end, we recorded time-resolved spectra of **G4** at pH3, where deprotonation of $(\text{G}^+)^\bullet$ is hindered.¹² This is shown in Figure 2b in the case of $(\text{TG}_4\text{T})_4/\text{Na}^+$:²⁸ the agreement with the monomeric radical spectrum is satisfactory in the 450-700 nm domain, not only regarding the spectral shape but also the extinction coefficients determined by dividing the observed transient absorption ΔA at 2.5 μs by $[e_{\text{hyd}}^-]_0$, which corresponds to that of generated electron holes. As predicted by quantum chemistry calculations, lower ϵ values were found for the band at 400 nm.^{26,28} Below this wavelength, it was impossible to determine reliable spectra due to strong photodegradation occurring in acidic solutions during the measurements.

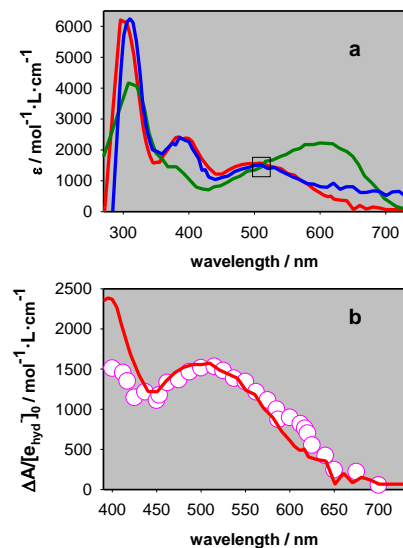


Figure 2. Reference monomeric **G** radicals. Absorption spectra of monomeric radical cation (\mathbf{G}^+) \bullet (red),¹² and deprotonated ($\mathbf{G}\text{-H1}$) \bullet radical (blue)¹² and ($\mathbf{G}\text{-H2}$) \bullet radicals (green),⁵² resulting, respectively, from loss of the protons at position 1 or 2 (Conspectus Figure); the black square indicates the spectral region where the ϵ values of all three radicals are similar ($\Delta\epsilon < 100 \text{ mol}^{-1}\cdot\text{L}\cdot\text{cm}^{-1}$).²⁷ (b) Transient absorption spectrum obtained for $(\mathbf{TG}_4\mathbf{T})_4/\mathbf{Na}^+$ in pH 3 at 2.5 μs (circles); ΔA is divided by $[\mathbf{e}_{\text{hyd}}^-]_0$, determined in the same experiment; the spectrum of the monomeric (\mathbf{G}^+) \bullet (red)²⁸ is represented with ϵ values.

In Figure 2a, we remark the existence of a quasi-isosbestic point around 500-515 nm. Thus, regardless the radical evolution in a given system, the total radical concentration at time t , $[\mathbf{R}]_t$, may be obtained from the ΔA value at this wavelength range. We found that for all the examined systems, $[\mathbf{R}]_t$ at 3 μs matches the concentration of hydrated ejected electrons, corresponding to the initially created electron holes ($[\mathbf{R}]_0 = [\mathbf{e}_{\text{hyd}}^-]_0$). This means that only a small fraction of the radical population, associated with our experimental error (5-10%, depending on the system), reacts at earlier times. Consequently, the survival probability of the total radical population is given by the decays recorded at the quasi-isosbestic point, setting ΔA at 3 μs equal to 1. The survival probabilities determined for **CT-DNA** and **OXY/K⁺** are shown in Figure 3.

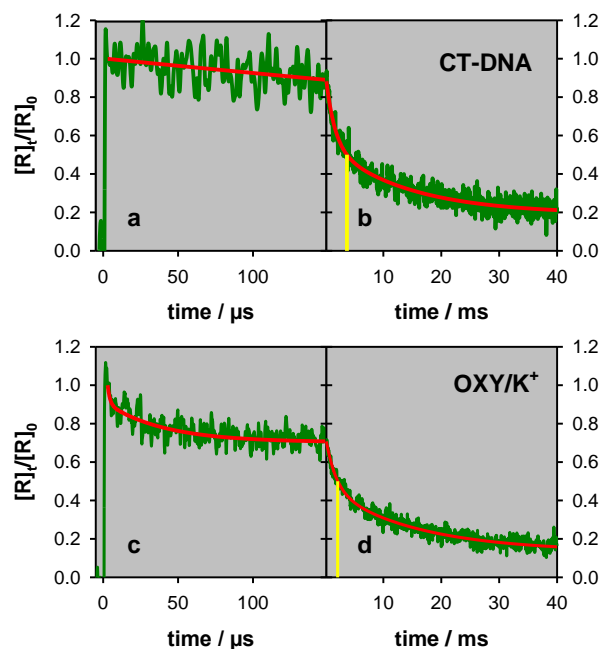


Figure 3. Survival probability of the total **G** radical population in **CT-DNA** (a and b) and **OXY/K⁺** (c and d). Experimental decays (green) recorded at 500 nm (a and b) and 512 nm (c and d) are fitted with multiexponential functions (red). The vertical yellow lines indicate $\tau_{1/2}$.

The radical half-lives $\tau_{1/2}$, that is the time at which $[R]_t/[R]_0 = 0.5$, are of a few milliseconds in both duplexes and **G4** (Table 1). The $\tau_{1/2}$ found for the duplex **S1•S2** (4 ms) is twice as long than those determined for the two parent single strands.²⁴ A similar increase in $\tau_{1/2}$ upon base-pairing is observed for adenine radicals in A-tracts.²¹ Larger conformational motions in single strands probably allow the systems to adopt more rapidly reactive conformations, increasing the reaction rate. Finally, we remark in Table 1 that $\tau_{1/2}$ of **G4** is much shorter in the presence of **K⁺** compared to **Na⁺**. The reasons underlying this effect are not clear yet.

CT-DNA. The transient absorption spectrum determined for **CT-DNA** at 50 μ s is shown in Figure 4a. It strongly resembles that of (**G-H1**)[•] radical reported for dGMP,¹² the agreement is quantitative, reproducing the ϵ values. However, as in the case of short duplexes,²⁴ the relative intensity at 700 nm in respect to that at 500 nm (~ 0.6) is higher than that characterizing the

monomer spectrum (~ 0.4). Such an enhanced absorption at the red part is predicted by quantum chemistry calculations for guanine radicals within G-C pairs.^{23,53} It was shown, in particular, that when the proton is transferred to the bulk water instead of cytosine, a weak intensity band appears around 650 nm.⁵³

We determined the deprotonation dynamics in **CT-DNA** by monitoring the transient absorption at 700 nm because only **(G-H1)**[•] absorbs at this wavelength (Figure 2a). To this end, we eliminated the much stronger absorption of e_{hyd}^- using efficient electron scavengers (N_2O and NO_3^-).³² Under these conditions, we observed a rise which is completed within $\sim 2 \mu\text{s}$ (Figure 4b); it can be approximated with a monoexponential function with a time constant of $0.3 \mu\text{s}$, in line with the fast deprotonation rate obtained for short duplexes using pulse radiolysis.⁵⁴ Subsequently, we found that the signal starts decaying slowly on the μs time-scale (Figure 4c), with a rate similar to that at 500 nm (Figure 3a). The indirect oxidation study reported instead a slow rise corresponding to a time constant of $7 \mu\text{s}$.⁵⁴ This discrepancy probably arises from anisotropic reaction kinetics between the oxidant and DNA bases in the duplex, as discussed in detail in reference 24.

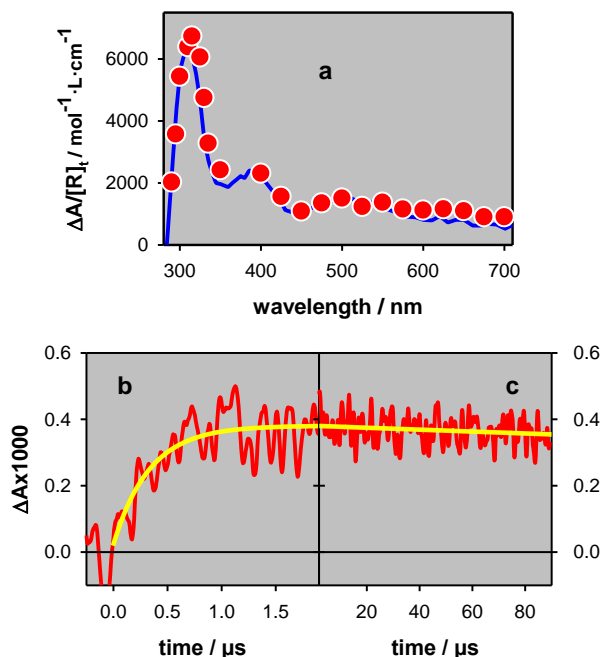


Figure 4. Guanine radicals in CT-DNA. (a) Transient absorption spectrum determined for aerated solutions at 50 μs (red circles); ΔA was divided by $[R]_t$. The blue line corresponds to the deprotonated radical of dGMP attributed to $(\text{G-H1})^\bullet$, represented with ϵ values.¹² (b and c) Transient absorption signals recorded for N_2O saturated solutions at 700 nm; yellow lines are derived from fits with monoexponential rise/decay; incident excitation intensity: $2.0 \times 10^6 \text{ W} \cdot \text{cm}^{-2}$.

The ϵ values of the $(\text{G-H1})^\bullet$ at 700 nm have not been determined experimentally for duplexes. However, the spectral shape in Figure 4a, in analogy with the computed spectra,⁵³ as well as the extent of rise in Figure 4b, strongly suggest that the quasi-entire population of $(\text{G}^+)^\bullet$ undergoes deprotonation within 2 μs .

Radicals in G4. Going from duplexes to four-stranded structures, the spectra recorded at 3 μs exhibit a broad absorption band in the visible, attributed to the coexistence of $(\text{G}^+)^\bullet$ and $(\text{G-H2})^\bullet$ radicals. As the hydrogen at position 1 of guanines (Conspectus Figure) is engaged in hydrogen bonding within the tetrads, deprotonation occurs at position 2. The existence of long-lived $(\text{G}^+)^\bullet$ in G4 and their deprotonation toward $(\text{G-H2})^\bullet$, was reported in a study performed via photosensitization.⁵⁵ In our studies, we quantified the population of these long-lived $(\text{G}^+)^\bullet$. To

this end, we reconstructed the time-resolved spectra with linear combinations of the spectra of monomeric radicals. An example is shown in Figure 5a, where the reconstruction of the transient absorption spectrum obtained for **OXY/K**⁺ at 3 μs is presented. The fraction of the radical population persisting as electron holes at 3 μs ranges from 0.25 to 0.50, depending on the **G4** structure.^{24,26-30}

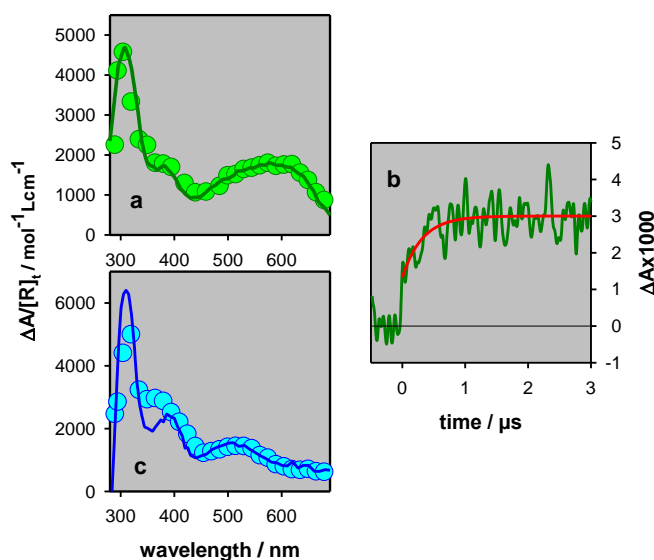


Figure 5. **G** radicals in **OXY/K**⁺. Transient absorption spectra (circles) at 3 μs (a, $[R]_t = [R]_0$) and 10 ms (c, $[R]_t = 0.3[R]_0$) determined for aerated solutions; the green line in (a) is a linear combination of the spectra corresponding to $(\text{G}^+)^{\bullet}$ (30%)¹² and the $(\text{G-H2})^{\bullet}$ (70%)⁵² radicals of monomeric guanosine; the blue line in (c) corresponds to the spectrum $(\text{G-H1})^{\bullet}$ of dGMP.¹² (b) Transient absorption trace at 620 nm recorded for N_2O saturated solutions (green); the red line corresponds to the fit with a monoexponential rise. Incident excitation intensity: $2.0 \times 10^6 \text{ W}\cdot\text{cm}^{-2}$.

The $(\text{G}^+)^{\bullet} \rightarrow (\text{G-H2})^{\bullet}$ deprotonation in **G4** spans several decades of time. It can be observed by the rise of the transient absorption signal at 600-620 nm, where the difference between the two radical spectra is particularly important (Figure 2a). The fastest part (Figure 5b) is probably related with outer tetrads that are most exposed to bulk water. Completion of the $(\text{G}^+)^{\bullet}$ deprotonation ranges from few decades of microseconds in tetramolecular **G4**²⁸⁻³⁰ to a few milliseconds in **TEL21/Na**⁺.²⁶ In most cases, $(\text{G}^+)^{\bullet} \rightarrow (\text{G-H2})^{\bullet}$ deprotonation is followed by $(\text{G-}$

$\text{H2}^\bullet \rightarrow (\text{G-H1})^\bullet$ tautomerization.^{24,26-28} An example is shown in Figure 5c, where the **OXY/K⁺** spectrum at 10 ms in the visible spectral domain corresponds to that of $(\text{G-H1})^\bullet$ in a concentration equal to 30% of $[\text{R}]_0$ (Figure 3d). Below 400 nm we observe the fingerprint of an unknown reaction intermediate, also detected for monomolecular **G4**.^{24,27} Tautomerization is not observed in **(TG4T)₄/K⁺**, where $(\text{G-H2})^\bullet$ radicals react rapidly through some other, currently unexplored, reaction path.²⁰

5. Correlation with final lesions

Although the present study deals with primary processes, our findings shed some light upon final lesions. For example, the formation of 8-oxodG, known to stem from hydration of $(\text{G}^+)^\bullet$, is in competition with the deprotonation process. Consequently, the level of 8-oxodG will be determined by the relative rates of these two reactions. This effect is illustrated by a comparative study on **TEL21/Na⁺** and **(TG₄T)₄/Na⁺**, for which 8-oxodG was analyzed by high performance liquid chromatography coupled to mass spectrometry following irradiation at 266 nm.^{26,28} The quantum yield for 8-oxodG formation ϕ_{ox} was found to be 3.2×10^{-4} for **TEL21/Na⁺** and lower than 10^{-5} for **(TG₄T)₄/Na⁺**. For the former system, in which part of the $(\text{G}^+)^\bullet$ population survives on the millisecond time-scale, ϕ_{ox} represents 7% of the photo-ionization quantum yield ϕ_1 . Deprotonation is much faster in **(TG₄T)₄/Na⁺**, being completed within 20 μs , thus rendering the reaction path leading to 8-oxodG less probable ($\phi_{\text{ox}}/\phi_1 < 0.003$).

As we mentioned above, we found that the largest part of the initially generated $(\text{G}^+)^\bullet$ population in the examined systems undergoes deprotonation. Therefore, we expect the major oxidative damage to arise from deprotonated radicals. As neither the radical concentration nor the decay patterns are affected by the buffer concentration (Figure S4a), we conclude that their

reactions do not involve buffer ingredients. Furthermore, no significant difference was found between the transient signals recorded for aerated and argon saturated solutions (Figure S4b). The latter observation rules out formation of imidazolone and oxazolone, known to be formed from (G-H1)[•] radicals exclusively in the presence of oxygen. In contrast, the formation of oxidized bases and strand breaks are not affected by oxygen.^{56,57} In the case of plasmid DNA, the yield of oxidized bases was found to be 20-fold higher than strand breaks.⁵⁶ Interestingly, strand breaks were detected in plasmid DNA following direct absorption of low-energy radiation,⁴⁸ but quantum yields were not determined.

Our studies, performed without external additives, show that deprotonated radicals react mostly on the millisecond time-scale in an “intrinsic” way. This means that G radicals disappear reacting with other parts of the nucleic acids and/or water molecules. Evidently, the reaction path may be altered by the presence of other chemical species in their vicinity, as happens in cells. The outcome will depend on the relative reaction rates of “extrinsic” and “intrinsic” paths, the final lesions being controlled by the faster process. The same reasoning is valid for the photo-ejected electrons which, in our experimental conditions appear to interact with genomic DNA, but may be scavenged by other chemical species present in biological conditions.

6. Conclusions and Outlook

The studies presented in this Account have challenged two widely accepted ideas regarding DNA damage and lead to important questions.

Firstly, we showed that direct absorption of low-energy UV radiation by DNA does generate guanine radicals. In the case of genomic DNA, the quantum yield of this process at 266 nm (2.0×10^{-3}) is similar to the sum of the quantum yields of all types of dimeric pyrimidine

photoproducts, both cyclobutane dimers and (6-4) adducts, following 254 nm irradiation (1.3×10^{-3}).⁵⁸ The generation of guanine radicals via direct photo-oxidation, occurring without the mediation of photosensitizers, has important repercussion on preventive measures: administration of antioxidants to protect DNA damage from low-energy UVB and UVA radiation, although proved efficient for indirect damage, cannot be operative for direct UV-induced radical generation.

Secondly, we demonstrated that 8-oxodG, a widely used oxidation marker, is not representative of the extent of oxidative damage when this is triggered by generation of radical cations. In this case, the major oxidative lesions should stem from deprotonated radicals and be context dependent, because of the long lifetime of these species. Photo-ionization quantum yields provide an estimation for the expected total amount of oxidative lesions. In association with analytical methods, they could contribute to quantify the various types of final reaction products and, possibly, lead to characterization of new ones. For example, the lesions resulting from (**G-H2**)[•] radicals, detected in all **G4** structures, remain to be identified.

To explain photo-ionization by low-energy UV photons, we proposed a complex path, encompassing recent knowledge of the electronic excited states in DNA. This hypothesis is supported by a series of observations associating photo-ionization with DNA secondary structure. It also explains the detection of 8-oxodG and strand breaks after UVB and UVA irradiation of DNA. A non-vertical mechanism is also suggested by electron detachment studies on DNA polyanions in the gas phase.⁵⁹ However, more effort is needed for its validation, fully characterizing the various steps that lead to electron photo-detachment and assessing the electronic, geometrical and dynamical factors that enter in play. We hope that this Account will

motivate theoretical studies which will explore this phenomenon in detail and provide a comprehensive picture.

An important question concerns the sites on which oxidative lesions are formed. This issue was tackled by a photo-sensitization study on telomeric **G4**.⁶⁰ It would be interesting to explore whether the reactive sites created by external oxidants are similar to those generated through direct UV-induced oxidation.

Finally, beyond the biological issues discussed above, the larger propensity of **G4** to undergo low-energy photo-ionization, compared to duplex DNA, deserves attention. **G4** structures appear appealing for applications in the field of nanotechnology.⁶¹ Their behavior as molecular wires, connected to external electric circuits, was reported.^{62,63} But so far, their properties as photoconductors have been not explored. In this respect, our results could have an impact on the design of optoelectronic devices inspired by four-stranded scaffolds, potentially containing other functional groups.

ASSOCIATED CONTENT

Supporting Information: Experimental Details; Saturation Effect; Additional Figures.

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Mr. Evangelos Balanikas graduated from the National Technical University of Athens (Greece) in 2017. He currently works as early stage researcher in the frame of the European Training Network “LighDyNAMics: DNA as a training platform for photodynamic processes in soft materials”. He prepares a PhD thesis on the UV-induced processes in **G**-quadruplexes.

Dr. Akos Banyasz obtained his PhD degree in Physical-Chemistry at the Eötvös Loránd University (Hungary) in 2006. He joined the CNRS in 2008 to investigate DNA photophysics and photochemistry induced by UV irradiation. He has expertise in time-resolved spectroscopy, from femtoseconds to milliseconds. Beyond DNA, his current research interests involve nonlinear spectroscopy and nonlinear 3D-printing.

Dr. Thierry Douki was trained in organic synthesis and joined the French Atomic Energy Commission in 1993. His research work is devoted to DNA damage and repair. He combines mechanistic studies on model systems with *in vitro* studies through development of specific analytical tools. The studied genotoxic agents include pollutants, oxidative stress and UV radiation.

Dr. Gérard Baldacchino obtained his PhD degree (1994) and Habilitation (2004) at the University Paris-Sud in the field of Molecular Chemistry. He joined the French Atomic Energy Commission in 1994, focusing his research on chemical effects induced by various types of radiation in aqueous systems. He developed sensitive transient absorption and fluorescence techniques to investigate complex radical mechanisms.

Dr. Dimitra Markovitsi received her PhD/Habilitation in Chemistry from the “Louis Pasteur” University (Strasbourg) in 1983. She joined the CNRS in 1981, where she is currently research director. She has expertise in photophysics and photochemistry, with emphasis on

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