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Coupling NMR to SANS: Addressing at once structure and dynamics in soft matter

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Abstract. In soft condensed matter, Small Angle Neutron Scattering (SANS) is a central tool to probe structures with characteristic sizes ranging from 1 to 100 nm. However, when used as a standalone technique, the dynamic properties of the sample are not accessible. Nuclear Magnetic Resonance (NMR) is a versatile technique which can easily probe dynamical information. Here, we report on the coupling of a low-field NMR system to a SANS instrument. We show that this original set-up makes it possible to obtain structural information and to simultaneously extract *in situ* on a same sample, long-range translational diffusion coefficient, T_1 and T_2 nuclear spin relaxation times. Such a feature is of major interest when a sample experiences a transient physical state or evolves rapidly. We illustrate the capabilities of alliancing these experimental methods by following the critical temperature-induced phase separation of a concentrated Poly(Methacrylic Acid) solution at its Lower Critical Solution Temperature. The characteristic size related to the domain growth of the polymer-rich phase of the gel is monitored by the evolution of the SANS spectra, while the dynamics of the sol phase (H_2O and polymer) is simultaneously characterized by NMR by measuring T_1 , T_2 and the diffusion coefficient. Great care has been taken to design a cell able to optimize the thermalization of the sample and in particular its equilibration time. Details are given on the sample cell specifically designed and manufactured for these experiments. The acquisition time needed to reach good signal-to-noise ratios, for both NMR and SANS, match: it is of the order of one hour. Altogether, we show that *in situ* low-field NMR/SANS coupling the NMR is meaningful and is a promising experimental approach.

Keywords: Critical phenomena, gelation, kinetics, LCST, PMAA, SANS, low-field one-sided NMR

1. Introduction

Gels are macromolecular networks. Describing their structure and physical properties comes to understand the processes leading to their formation. Gelation is the result of a mechanism where individual molecules (monomers or polymer chains) are able to bind to at least three of their neighbors. The population of the remaining free

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molecules is soluble (hence the term of sol) until the appearance of a structure of infinite size: the gel. From a fundamental point of view, gelation is not a thermodynamic transition, but instead a critical transition of connectivity. Regardless of the local rules governing this connectivity, the machinery of critical phenomena can be used to describe all physical, chemical or biological processes where association of elementary molecules, initially separated from each other, leads to the formation of large clusters, then to macroscopic phases. The measurable static and/or dynamic quantities diverge at the gelation threshold and follow a power law behavior when they are expressed as a function of the gap to this point [1,2].

Catching simultaneously sharply evolving dynamical and structural quantities demands experimental techniques able to monitor the spatial dependence of correlation times. Light [3] and neutron scattering, in particular Neutron Spin-Echo (NSE) [4], have indeed shown large success in the field. But, despite the scientific need, the possibility to reach the 0.1 to 10^{-2} nm^{-1} Q range, unreachable with light scattering, is at the limit of the current NSE spectrometers. This is the nominal Q range of most of the SANS instruments but, with this technique, when used in standalone, the dynamical properties of the samples are not accessible. This kind of information can be easily obtained with Nuclear Magnetic Resonance (NMR) techniques [5]. The accessible time-scales in NSE are from few picoseconds up to tens of nanoseconds (exceptionally few hundred of nanoseconds on specific instruments) while, in NMR, correlation times go down to picoseconds in relaxation experiments and up to tens of seconds for diffusion experiments. Coupling an NMR system to a SANS instrument therefore seems an appealing solution to fill the current $[Q, t]$ gap existing among the neutron high-resolution quasi-elastic techniques.

In this paper we report the first, to our knowledge, coupling of NMR and SANS and address the temperature-induced phase separation of a concentrated Poly(Methacrylic Acid) (PMAA, $[-\text{CH}_2-\text{C}(\text{COOH})(\text{CH}_3)-]_n$) solution at its Lower Critical Solution Temperature (LCST). The driving force of such transitions is usually the entropy gain associated to the release of the solvent: as the water molecules experience a reduced number of hydrogen bonds with the polymer chains, their entropy increases. The conformation of this polyelectrolyte is driven by both the intra- and inter-chain competitions of *i*) intermolecular hydrogen bonds between carboxyl groups, and *ii*) hydrophobic interactions induced by the presence of methyl side-chains. It may be tuned by the pH and/or ionic strength. At low pH when PMAA stands in its acidic form (degree of ionization ~ 0), concentrated solutions have been reported to form thermo-reversible gels [6]. A gelation process has been confirmed by rheological measurements and simple transmittance experiments [6–9]: at low temperatures the solution is transparent whereas it turns opalescent above a so-called *cloud-point* temperature at $T_c \approx 60^\circ\text{C}$. Usually, for polymer solutions showing a LCST, while the macroscopic state of the solution seems to remain unchanged above the cloud-point, the nanoscale structure state continues to evolve, especially in the semi-diluted regime [10].

Up to date, this temperature-induced phase separation of PMAA has been poorly addressed in the literature. It has been proposed that the gelation process could be induced by physical cross-linking of neighboring chains. If this is the case, one expects that the structuration of the gel has a strong influence on the behavior of the sol phase (the solvent). In this work, the characteristic sizes related to the domain growth of the polymer-rich phase are monitored by the evolution of the SANS spectra while the dynamics of the protonated moieties of the system (H_2O and polymer) are simultaneously characterized (T_1 , T_2 and diffusion coefficients) by NMR methods.

2. Materials and methods

2.1. Sample

To minimize the incoherent background and to optimize the coherent contrast between polymer and solvent, SANS experiments on aqueous polymer solutions are usually performed using D_2O . In the present framework of NMR/SANS coupling, to maximize the NMR sensitivity, the NMR radiofrequency of the probe was set on the resonance of the ^1H nuclei Larmor frequency. A compromise had therefore to be found on the isotopic composition of the solution and we have used a $\text{H}_2\text{O}/\text{D}_2\text{O}$ mass ratio of 50%. With this isotopic mixing, the PMAA LCST gelation transition occurs at *ca.* 65°C .

PMAA (fully hydrogenated) was purchased from Polymer Science Inc. Its molar mass and molar mass distribution were obtained using Size Exclusion Chromatography (SEC). Prior to analysis, the polymer was modified by methylation of the carboxylic acid group using trimethylsilyldiazomethane. The samples were analyzed in tetrahydrofuran (THF) at a concentration of 10 mg/ml at 25°C. The column used was a polystyrene mixed C13 column for organic solvents. The setup was equipped with a refractive index (*RI*) detector at 658 nm and a light scattering detector. The average molar number mass $M_n = 313 \text{ kg.mol}^{-1}$, weight mass $M_w = 391 \text{ kg.mol}^{-1}$ and the polydispersity ($\text{PDI} = 1.25$) value were derived from the *RI* signal using a calibration curve obtained with poly(methyl methacrylate) standards purchased from Polymer Laboratories. PMAA was dissolved, at room temperature, at 80 g.l^{-1} in water ($\text{H}_2\text{O}/\text{D}_2\text{O}$ mass ratio of 50%) two days before the experiment. From viscosimetric measurements [11], we estimate that $C^* \approx 0.26 \text{ g/l}$.

2.2. SANS instrumental settings

In order to ensure the exact same experimental conditions for both the SANS and the NMR measurements, a home-made single-sided NMR spectrometer (see details in Section 3) was installed on the PAXY instrument [12] of the LLB (Laboratoire Léon Brillouin, Saclay, France). The SANS incident wavelength and sample to detector distances were, $\lambda_0 = 5.0 \text{ \AA}$ and $D_{\text{SD}} = 2.5 \text{ m}$ respectively. The beam was collimated by 2 circular aperture slits of 16.0 mm and 7.6 mm, the second one being placed immediately before the sample. The surface of the neutron beam enlightening the sample was 58 mm^2 . At each temperature, the SANS spectra were measured for 90 minutes.

3. NMR set-up

3.1. A home-made portable single-sided NMR setup

Many single sided NMR systems have been proposed in the literature [13]. Our approach to magnet design is based on the Spherical Harmonic Decomposition methodology to lead to a 12-fold cylindrically symmetric one-sided constant-gradient magnet without pole pieces [14,15], which has been used already to measure one-dimensional proton density profiles as well as relaxations and diffusions [16,17]. Our permanent magnet is a cylinder of 24 cm in diameter, 22 cm in height containing NdFeB magnets and weights $\sim 40 \text{ kg}$.

Its design consist in a simple dipole pointing along the *z* direction in the laboratory frame [14,15]. To ease its construction, we assembled and glued all magnet pieces non-magnetized and we magnetized it afterwards in a one-step process using a powerful electromagnet at the LNCMI-CNRS facility [14].

This device produces a magnetic field strength of $\sim 0.3 \text{ T}$ at 2 cm far from its upper surface which has a linear magnetic field gradient $g = 3.43 \text{ T/m}$ [17]. A photo of the magnet as well as a field mapping above its surface can be seen on Fig. 1. Because of the absence of cryogenic fluids and due to its compactness and sample accessibility, it is an ideal candidate for multimodal studies.

A single channel Tecmag's Scout spectrometer and a Tomco[®] BT01000-Gamma power amplifier were used. The radio-frequency probe consisted of a rectangular shape copper coil with 10 turns tightly wound to fit a 1 mm thick Hellma[®] quartz cuvette, with total length 50 mm, and an external box in which the parallel and series mounted capacitors to the coil were accommodated, for the tuning and matching of the probe.

3.2. NMR pulse sequence

The longitudinal T_1 , transverse T_2 relaxation times and the self-diffusion coefficient, D_t were successively measured as a function of the sample temperature, at the ^1H Larmor frequency of 13.61 MHz. At this frequency, by discriminating all the other elements/isotopes of the system, only information about the PMAA and H_2O protons were accessible.

T_1 was measured by inversion-recovery, T_2 using a Carr–Purcell–Meiboom–Gill (CPMG) sequence and D_t by a Hahn echo sequence (Fig. 2). In the diffusion and T_1 measurements, CPMG-like acquisition was used to

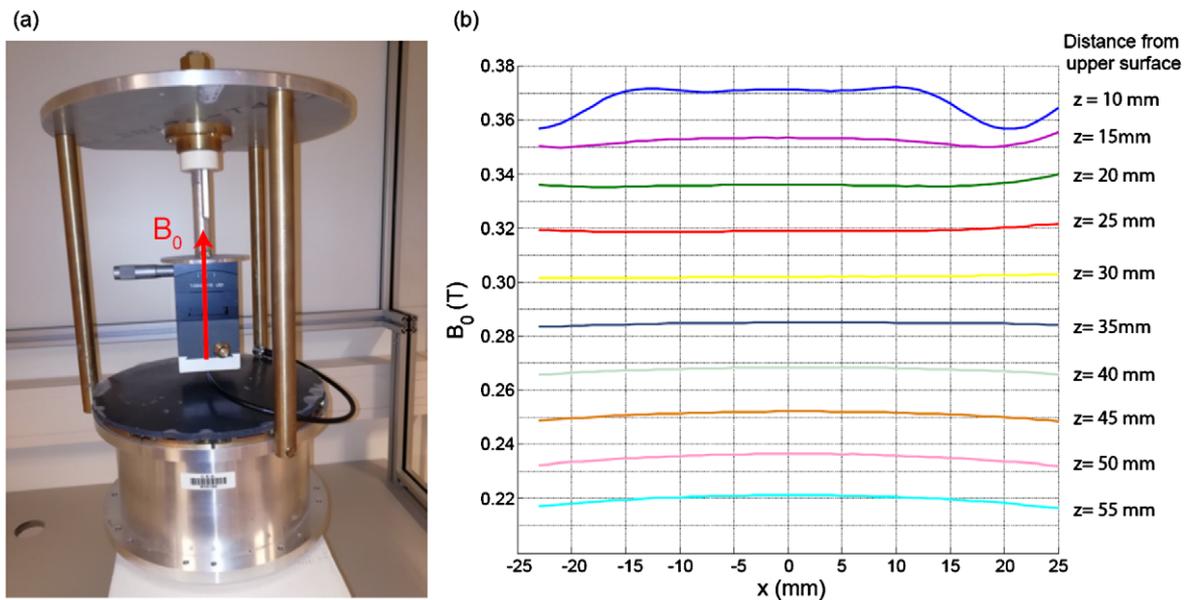


Fig. 1. (a) Setup of NMR magnet and probe: the cylindrical NdFeB magnet assembly sits on an aluminum table and the goniometric probe is located along the central axis above the magnet. The sample is fixed inside the coil of the probe, which is precisely positioned by two non-magnetic goniometric stages fixed perpendicular to each other. The vertical position of the probe is controlled by a central threaded rod, which is fixed on an aluminum disk at 28 cm from the top surface of the magnet. (b) Magnetic field profile measured above the surface of the magnet in the XZ plane using a 3-axis hall sensor from senis AG (model 3MH3-0.1%-2T). Ten equidistant vertical positions were scanned and from the figure we can clearly identify the linear field gradient region of the magnet located above the center of the magnet ($x = 0$) at $z = 30$ mm from its upper surface.

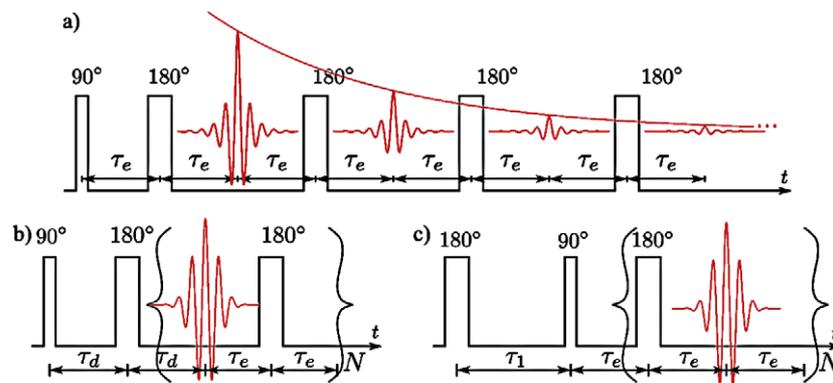


Fig. 2. Pulse sequences used to measure: (a) the transverse relaxation times T_2 , (b) the self-diffusion coefficients, D_f , and (c) the longitudinal relaxation times T_1 . The time delays are: τ_e the echo-time, τ_d , the diffusion time and τ_1 the recovery time. CPMG was used to increase the signal sensitivity during detection and a large number of echoes ($N = 20$) were summed together to form the signal intensity.

increase the signal intensity and the 20 first echoes were added together for each scan. For the inversion recovery experiment, 16 recovery times were logarithmically set from 0.1 to 10,000 ms. The diffusion times τ_d , were set to: 0.464 ms, 0.563 ms, 0.663 ms, 0.762 ms, 0.861 ms, 0.960 ms, 1.060 ms and 1.159 ms. The echo time $\tau_e = 50 \mu\text{s}$. The duration of the 90 and 180° radiofrequency pulses were equal to 0.5 μs and 1.0 μs , respectively, and the repetition time was set to 10 s for all experiments.

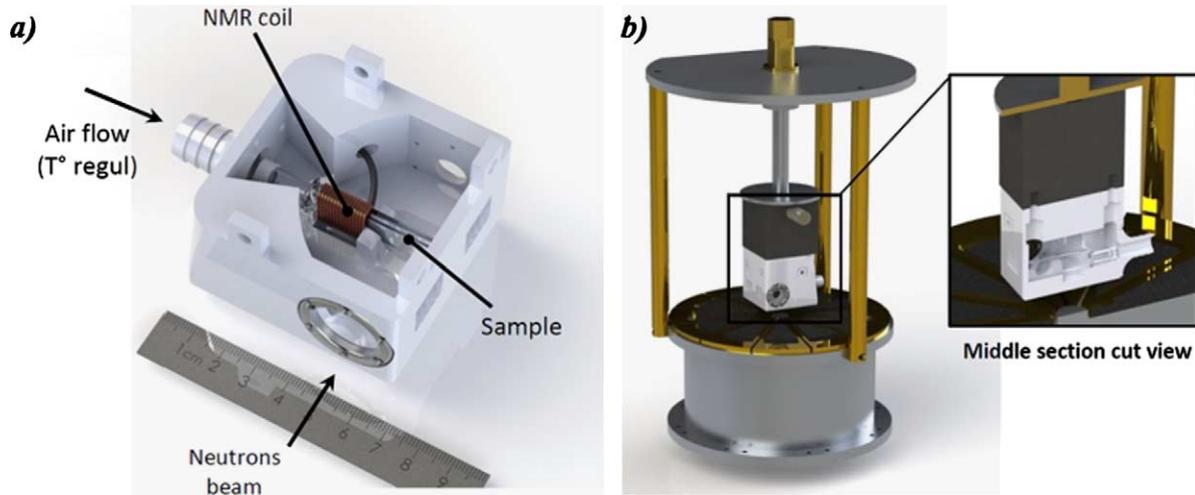


Fig. 3. (a) picture of the NMR-neutron probehead (cover not shown). The copper NMR coil tightly surrounds a part of the Hellma® sample cell at immediate proximity of the neutron beam section. (b) 3D view of the probehead adaptation onto the NMR spectrometer. Here, the NMR-neutron probehead cover necessary to control the heated air stream used for the sample temperature regulation is shown.

4. A probehead for concurrent SANS and NMR measurements

As the sample cell is a critical parameter when it comes to study temperature driven critical phenomena (like the PMAA gelation process we are addressing in this study), great care has been taken to design a cell able to optimize the thermalization of the sample and in particular its equilibration time. In this section, we provide details on the specific sample cell manufactured for this work.

4.1. Design of a cell for in-situ NMR measurements on a SANS machine

To stay in tune with the classical soft matter conditions used on SANS spectrometer, the NMR-Neutron probehead has been designed to accommodate a classical $50.0 \times 12.6 \times 1.0 \text{ mm}^3$ (height \times width \times thickness) Hellma® quartz cuvette able to offer a total neutron beam cross-section of the order of 1 cm^2 .

This Neutron-NMR probehead is shown on Fig. 3. It is made from PLA (polylactic acid) by 3D printing. It is closed to ensure the thermalization of the sample by a stream of heated air generated by a heat-gun and driven to the cell entrance by a standard plastic hose (see entrance funnel on Fig. 4). The neutron beam passes through two quartz windows. This geometry offers the large detector an angular access of 40° required for SANS measurements.

4.2. Optimization of the NMR-neutron probehead thermalization

To thermalize the sample, electric heating devices are usually used. Here, in order to avoid any magnetic field disturbances induced by electric current, we have chosen to use a stream of heated gas. The air stream within the NMR-Neutron cell has been simulated using the COMSOL® Multiphysics software. The simulations show that the air flow-rate is able to thermalize the sample (Fig. 4(a)). Experimentally, the sample temperature was directly measured inside the measuring cell using an optic fiber temperature sensor (OTG-M420, OpSens®) placed outside of the neutron beam cross-section. Starting from room temperature, when passing the set-point to 70°C , the required temperature is reached after 10 minutes and a stability within 0.1°C can be maintained for hours. We have checked that the sample temperature is not affected by overnight changes of ambient temperature in the neutron guide hall.

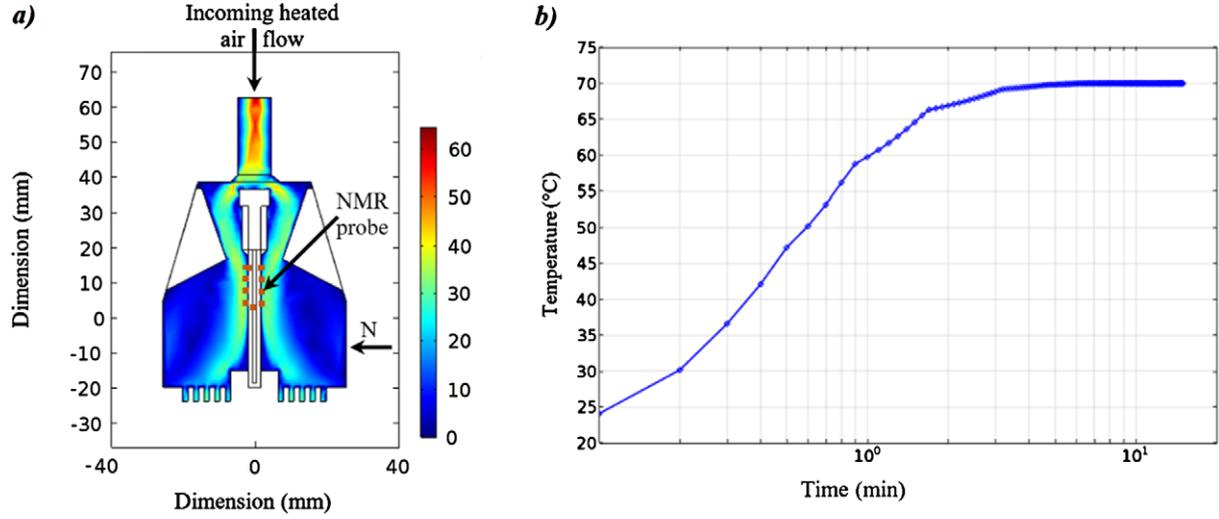


Fig. 4. a) Simulation of the air-flow within the NMR-neutron probehead (sectional and top view). The color code (in m.s^{-1}) is shown on the right. The incoming heated air flow, the position of the NMR probe-head and the axis of the neutron beam (N) are shown. b) Experimental time-dependence and stability of the ambient temperature within the NMR-neutron sample during a temperature jump of 50°C .

5. Coupling of the one-sided NMR system to the SANS machine

The adaptation of the one-sided NMR spectrometer on the SANS machine was rather straightforward. The major modification to the regular PAXY instrument set-up was the removal of the sample changer and of the goniometer usually installed underneath. This was necessary to minimize the interaction of the NMR magnet with such massive magnetic materials. To recover the proper position of the sample in the beam, these parts are replaced by pieces of high-density polyethylene. The hybrid NMR-SANS set-up is shown on Fig. 5.

6. Results

6.1. SANS

Small angle neutron scattering measurements have been performed at increasing temperature below and above the cloud point. At each temperature change, successive SANS spectra were measured for 5 minutes each, until no evolution of the scattering was detected. The equilibration of the sample was of the order of 30 minutes. The SANS data are shown on Fig. 6.

At low temperature, below the cloud-point, PMAA solutions behave as classical semi-diluted solutions. As shown on Fig. 6, the scattering intensity of PMAA solution at 80 g.l^{-1} can be fitted with a classical Ornstein-Zernike function:

$$I(q) = (\rho_{\text{PMAA}} - \rho_{\text{water}})^2 I_{\text{OZ}}(q=0) / (1 + q^2 \xi^2) \quad (1)$$

where $I_{\text{OZ}}(q=0)$ is a prefactor, ρ_{PMAA} and ρ_{water} are the Neutron Scattering Length Densities (NSLD) of PMAA and of the solvent, ξ corresponds to the correlation length of the semi-diluted solution.

Above the cloud-point, a strong intensity is detected at $q < 0.01 \text{ \AA}^{-1}$, indicating the formation of a large-scale structure. Classically, in the field of gelation, this corresponds to a spinodal decomposition of the system where a phase with a high polymer concentration coexists with a low polymer concentration one. The creation of a large

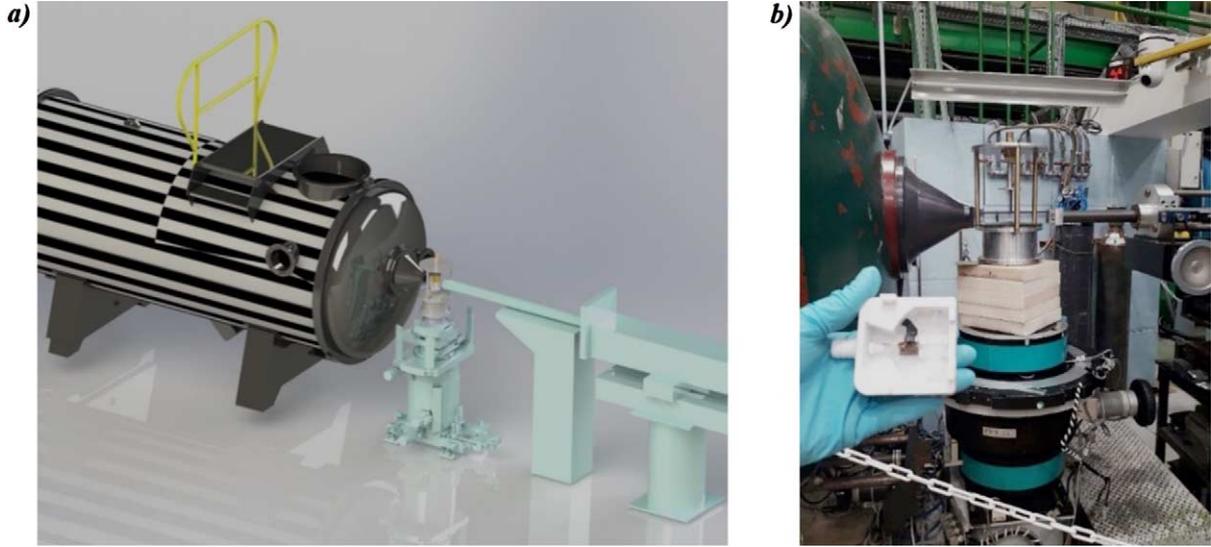


Fig. 5. a) 3D model of the LLB SANS instrument PAXY. From right to left, the neutron guide (in turquoise blue), the one sided-NMR spectrometer (at the center of the picture on a) and b)) and the neutron detector box (zebra) are shown. b) Real picture of the central part modeled in a). The NMR magnet is seen in the middle of the picture and the NMR-neutron probehead is shown at the forefront. The copper NMR coil surrounding the Hellma[®] cell is set in place. The NMR console (not shown) was 2 meters away from the sample position.

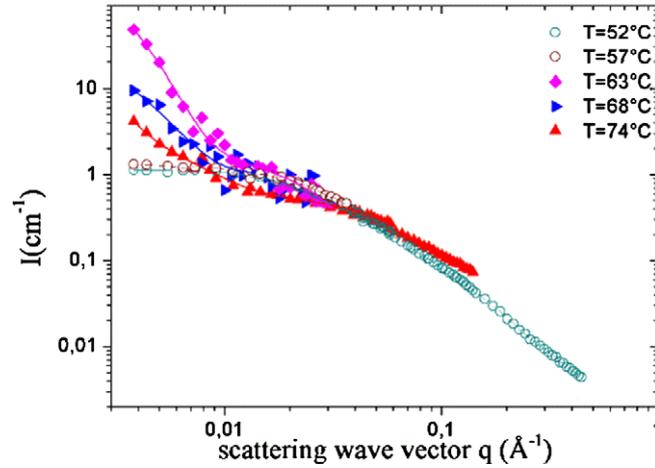


Fig. 6. Scattering intensity of PMAA solution at 80 g.l^{-1} at different temperatures below and above the cloud-point temperature ($T_c \approx 60^\circ\text{C}$). Fits are shown as full lines. At 52 and 57°C , *i.e.* below t_c , the data follow the pure Ornstein–Zernike function Eq. (1). Above T_c , at 63, 68 and 74°C , to account for an additive Porod contribution at $q < 0.01 \text{ \AA}^{-1}$ (see text), the data are fitted according to Eq. (4).

interface separating these two phases induces a q^{-4} Porod contribution. The total intensity above the cloud-point temperature can then be accounted for by:

$$I(q) = \frac{(\rho_{\text{PMAA}} - \rho_{\text{water}})^2 I_{\text{OZ}}(q=0)}{1 + q^2 \xi^2} + \Phi_{\text{conc}}(1 - \Phi_{\text{conc}}) \frac{2\pi(\rho_{\text{conc}} - \rho_{\text{diluted}})^2}{q^4} \cdot \frac{S}{V} \quad (2)$$

where S/V stands for the interface area per volume of the highly concentrated phase, Φ_{conc} for its volume fraction, ρ_{conc} and ρ_{diluted} for the NSLD of the concentrated and diluted phases respectively. These latter quantities are a

Table 1

Interface area per volume of the 80 g.l⁻¹ PMAA solution as a function of temperature. These values are obtained by a fit of data shown on Fig. 6 with Eq. (4)

T (°C)	52°C	57°C	63°C	68°C	74°C
S/V (mm ⁻¹)			87	65	8

function of $\varphi_{\text{PMAA_conc}}$ and $\varphi_{\text{PMAA_dil}}$ the volume fractions of PMAA within the concentrated and diluted phases respectively:

$$\begin{aligned}\rho_{\text{conc}} &= \varphi_{\text{PMAA_conc}}\rho_{\text{PMAA}} + (1 - \varphi_{\text{PMAA_conc}})\rho_{\text{water}} \\ \rho_{\text{diluted}} &= \varphi_{\text{PMAA_dil}}\rho_{\text{PMAA}} + (1 - \varphi_{\text{PMAA_dil}})\rho_{\text{water}}\end{aligned}\quad (3)$$

After the phase transition, $\varphi_{\text{PMAA_conc}} \gg \varphi_{\text{PMAA_dil}}$ and the NSLD of the diluted phase is therefore very close to the one of water $\rho_{\text{diluted}} \sim \rho_{\text{water}}$. This is a classical situation in the field of LCST studies [10]. The contrast term $(\rho_{\text{conc}} - \rho_{\text{diluted}})^2$ becomes $\varphi_{\text{PMAA_conc}}^2(\rho_{\text{PMAA}} - \rho_{\text{water}})^2$, and the Eq. (2) then reduces to:

$$I(q) = \frac{(\rho_{\text{PMAA}} - \rho_{\text{water}})^2 I_{\text{OZ}}(q=0)}{1 + q^2 \xi^2} + \Phi_{\text{conc}}(1 - \Phi_{\text{conc}}) \frac{2\pi \varphi_{\text{PMAA_conc}}^2 (\rho_{\text{PMAA}} - \rho_{\text{water}})^2}{q^4} \cdot \frac{S}{V} \quad (4)$$

As clearly seen on Fig. 6, the q^{-4} Porod contribution below $q < 0.01 \text{ \AA}^{-1}$ shifts to smaller q when the temperature increases. This evidences that above the LCST transition the aggregates within the highly concentrated polymer phase grow. As a consequence, their surface over volume ratio decreases.

As the Ornstein–Zernike contribution remains very similar on the whole temperature range, we can consider to a good approximation that at the local scale the organization of the polymer chains is almost unaffected by the onset of the LCST transition.

The Ornstein–Zernike contribution is fitted at low temperature and is kept constant over the whole temperature range. The Eq. (4) then fits nicely the data on Fig. 6. Provided that upon an increase of temperature, Φ_{conc} and $\varphi_{\text{PMAA_conc}}$ vary slightly compared to the growth of the domain size, it shows that the interface area per volume of the highly concentrated phase has been reduced by almost one order of magnitude from 63°C to 74°C (Table 1).

6.2. NMR

6.2.1. Relaxation times T_1 and T_2

Figure 7 shows the curves obtained with the inversion-recovery experiments for different temperatures. In order to obtain the value of T_1 , we applied bi-exponential fit of the experimental data:

$$f_1(t) = 1 - (a.e^{-t/T_{1a}} + b.e^{-t/T_{1b}}) \quad (5)$$

where a and b are the intensities of the two contributions with relaxation times T_{1a} and T_{1b} , respectively. The temperature dependence of these two longitudinal relaxation times, a and b , are shown on Fig. 7.

The transverse relaxation T_2 decay curves for different temperatures are shown on Fig. 8. Two transverse relaxation times were derived from a bi-exponential model fitting:

$$f_2(t) = a.e^{e(-t/T_{2a})-t/T_{2a}} + b.e^{-t/T_{2b}} \quad (6)$$

where a and b are the amplitudes of the relaxation components with transverse relaxation times T_{2a} and T_{2b} respectively.

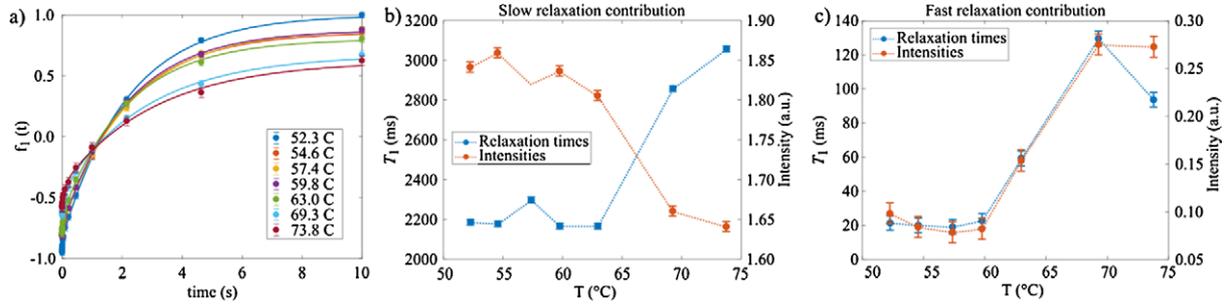


Fig. 7. NMR T_1 experiments: a) Inversion recovery curves for the different temperatures. b) and c) Temperature dependence of the relaxation times of the two components of T_1 , according to the data fitting with a bi-exponential model Eq. (5). The integrated intensities of these two relaxation time contributions are also shown.

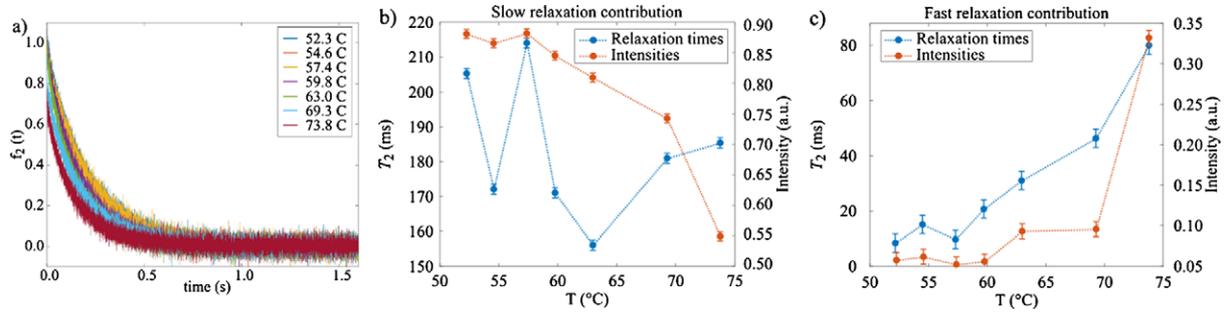


Fig. 8. NMR T_2 experiments: a) T_2 decay curves as a function of temperature, obtained from CPMG experiments. b) and c) Temperature dependence of the relaxation times of the two components of T_2 , according to the data fitting with a bi-exponential model Eq. (6). The integrated intensities of these two relaxation time contributions are also shown.

6.2.2. Self-diffusion coefficient experiments

The results of the diffusion experiments as a function of temperature are shown on Fig. 9. These curves are determined by the intensities of the signal for each diffusion time $S(t)$, recorded by applying the sequence shown on Fig. 2(a). The theoretical model writes:

$$S(t) = \log(s(\tau_d)/s(\tau_0)) \quad (7)$$

where $s(\tau_d)$ are the intensities of the signal for the different diffusion times τ_d . $s(\tau_0)$ is the intensity of the signal of the first diffusion time of each experiment for the diverse temperature. The diffusion coefficients related to the diffusion times throughout the function [13]:

$$s(\tau_d) = s(\tau_0) \exp(-2\tau_d/T_2 - 2\gamma^2 Dg^2 \tau_d^3/3) \quad (8)$$

where γ is the ^1H gyromagnetic ratio, g is the magnetic field gradient and T_2 is the transverse relaxation time from the Hahn echo experiments (see Fig. 2(b)). This relation can be re-written as:

$$\log(s(\tau_d)/s(\tau_0)) = \frac{-2}{T_2} \tau_d - \frac{2}{3} \gamma^2 Dg^2 \tau_d^3 \quad (9)$$

thus, D_t can be determined by the simple data fitting with a function:

$$f(x) = Ax + Bx^3 \quad (10)$$

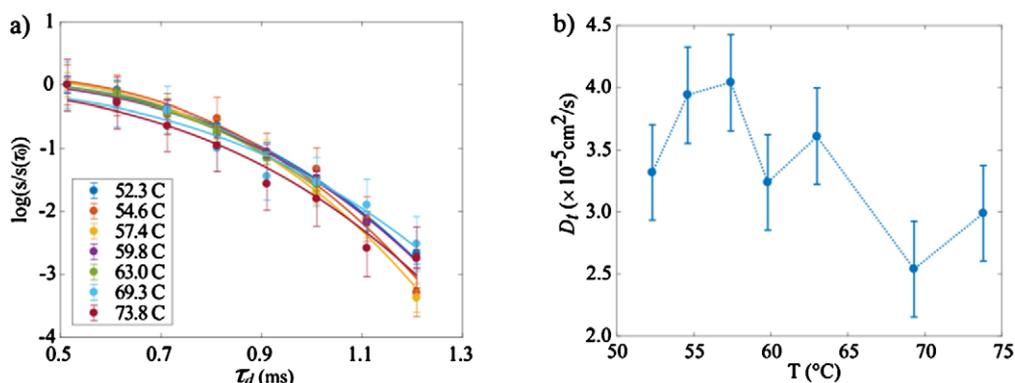


Fig. 9. a) Diffusion curves obtained using the Hahn echo sequence as a function of the temperature. b) Self-diffusion coefficients of the protons of the system, determined by data fitting of data shown in a) according to Eq. (10).

where $A = -2/T_2$ and $B = 2\gamma^2 Dg^2/3$.

7. Discussion

Using the bimodal NMR-SANS equipment described above, along with a regular SANS scattering instrument, we have been able to measure the common low field NMR parameters: T_1 , T_2 and the translational diffusion coefficient.

The gelation process we have been focusing on, is detected at the expected temperature of 65°C by SANS. Concomitant NMR measurements show the clear phase transition in the same temperature region. More specifically, abrupt changes of both T_1 relaxations times are observed at this threshold temperature. The diffusion coefficient measured in these experiments may be compared with values in pure water [18] *i.e.* $3.9 \times 10^{-5} \text{ cm}^2 \text{ s}^{-1}$ at 50°C and $5.6 \times 10^{-5} \text{ cm}^2 \text{ s}^{-1}$ at 70°C . In the liquid/low temperature state, the diffusion coefficient for water in the PMAA solution is slightly lower, as expected and reflected by the viscosity of the solution. The water diffusion coefficient is rather constant up to the cloud point and then decreases down to $2.5 \times 10^{-5} \text{ cm}^2 \text{ s}^{-1}$ at 75°C . This unusual behavior reflects either a strong interaction of the water inside the 3D network of the gel, or a restricted motion inside the polymer mesh.

Nevertheless, a thorough interpretation of the NMR data is challenging. First, here, the NMR signal is a composite information: both the protons from the solvent and from the PMAA contribute to the observed signal. Work is underway to find a way to better discriminate these two contributions. For future studies, a possible way out will be the use of fully deuterated water or PMAA.

At the present stage of development, we have made the choice *i)* to avoid any neutron beam-induced activation of the metallic (copper) NMR coil, *ii)* suppress any parasitic SANS signal induced by the scattering of this coil and *iii)* eliminate any reproducibility issues due to a change of the coil position in the beam upon sample change. As a consequence, while they have been measured simultaneously and on the very same sample, the signal recorded by NMR and SANS were coming from slightly different volumes of the sample.

During the experiment, when increasing the sample temperature to reach the LCST region, we have noticed that the fraction of the sample embraced by the radiofrequency signal generated by the NMR coil, was showing critical opalescence few degrees before the expected gelation temperature. The fraction of the sample few millimetres away from this milky spot was actually perfectly transparent.

In the present study of a critical gelation phenomena, this localised opalescence can with not doubt be assigned only to a local increase of the temperature: the opalescent fraction of the sample has exceeded the LCST temperature while the transparent fraction remained below.

We therefore conclude that, even with a recyclable delay of 10 s for the NMR sequences pulses (0.16 s in the CPMG experiment and $\sim 20 \mu\text{s}$ for inversion recovery and Hahn echo experiments), the dissipation of the $\sim 250 \text{ W}$ radiofrequency power was still enough to induce a noticeable local heating of the sample. We estimate this local temperature increase to be of the order of a few centigrades.

To overcome local heating, so as to reach a uniform temperature over the whole sample volume, instead of measuring a steady sample enclosed in the cell, we could create a mild flow of the sample by circulating, in the cell, the sample from (and to) a mother reservoir. This microfluidic-like strategy seems an easy way to lift the issue of NMR coiled induced inhomogeneous heating of the sample, but is only adapted to liquids and/or low viscosity samples.

For solid or gels samples, a direct way to overcome the heating issue will be to position the NMR coil directly in the neutron beam. Such a setting takes advantage that in the SANS regime, most of the metallic materials show a moderate (but Q dependant) scattering. As copper activates with neutrons, a good coil material candidate would be aluminium (^{27}Al): its radioactive half-life being of 2 minutes, such a coil would have fully deactivated after a duration of the order of 20 minutes after the closure of the neutron beam.

In the future, the use of advanced NMR techniques, such as T_1-T_2 , T_2-T_2 and $D-T_2$ bi-dimensional experiments [13] could be implemented and give more detailed insight on the physics (structure and dynamics) of the sample.

8. Conclusion

In this paper, we have demonstrated that the coupling of a single-sided NMR system with a SANS instrument is experimentally possible. For this purpose, a home-made portable single-sided NMR setup was properly adapted to the SANS machine, and bimodal experiments were simultaneously carried out. The acquisition times needed to reach good signal to noise ratios, of the order of one hour for both NMR and SANS, match. Altogether, we show that the *in-situ* low-field NMR/SANS coupling the NMR is meaningful and is a promising experimental approach.

As both structural ($S(Q)$) and dynamical information (T_1 , T_2 , diffusion coefficients) can be derived by a single experimental procedure, this hybrid technique could significantly contribute to the understanding of the microscopic phenomena at play in condensed and soft matter that define macroscopic properties.

Such an *in-situ* SANS/NMR coupling experimental strategy is relevant in the field of chemical physics to finely probe systems like polymers, membranes, colloids in scientific fields spanning from geology (water management in soils, oil industry, etc.), electrochemistry (structural and related dynamical behaviours of electrolyte upon additives) to biophysics.

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