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Single-cell bacterium identification with a SOI optical microcavity

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Photonic crystals and microcavities can act as on-chip nano-optical tweezers for identification or manipulation of biological objects. So far, virus and bacteria optical trapping has been achieved, their presence in the vicinity of the optical resonator being deduced from a shift of the resonant wavelength. Here we show that bacteria can not only be trapped but identified as well if the trapping time is long enough. A silicon on insulator microcavity with a properly tuned quality factor allows to achieve enhanced trapping times. By combining spatial and temporal observation of bacteria – cavity interaction, the optical identification of three different kinds of bacteria is demonstrated.

Biological objects are strategic components for medical and pharmaceutical development, faster clinical diagnosis and food processing control. The field of photonics has proven to be extremely suitable for the study of these objects. The first tool that allowed their trapping and handling is optical tweezers,^{1,2} based on the concentration of a laser beam through a microscope objective. Unfortunately, this method is limited by the macroscopic scale of the implemented set-up and especially by the damage caused on cells^{3,4} and bacteria⁵⁻⁷ during optical manipulation. On the contrary, the evanescent field of on-chip optical resonators offers a low light intensity energy that could minimize biological object damage. Therefore, they are more suitable for biomolecules,⁸⁻¹² virus^{13,14} and bacteria^{15,16} manipulation, at a microscopic scale. For the past 20 years many designs of optical resonators have emerged. From the 1D^{17,18} and 2D¹⁹⁻²¹ photonic crystals to toroid,²²⁻²⁴ ring,²⁵ cylindrical²⁶ and microspherical^{27,28} resonators, extremely narrow resonances and quality factors as high as 10^9 have been reached. With their integration in optofluidic devices, these structures allow micro and nano objects manipulation²⁹⁻³⁵ and biological object detection.^{37,38} More specifically, at the cell scale, despite the low index contrast with liquid medium, it has been proven that bacteria trapping is achievable on H-L types PhCs microcavities.¹⁵ For these structures, the trap stiffness was quantified and the confined Brownian motion of the bacteria has revealed power dependence to the light power excitation.¹⁵ In this direction, Lin *et al.* have demonstrated that this optical trap do not affect *Escherichia coli* viability by studying its photothermal damage. Thus, according to the theory, designed lineic microcavities appear to be suitable tools to manipulate bacteria.

Most of literature work has been focused on detection where high quality factor structures are required to precisely detect the shift of the resonance wavelength due to refractive index variation near the resonator^{12,36,37} (Figure 1(a)). What we propose here is the use of a broader resonance, adapted for single bacterium analysis and label-free identification (Figure 1(b)). To achieve this condition, the resonance must be wide enough to have an overlap between the resonance without bacterium and the resonance when the bacterium interacts with the field in the cavity (redshift).

Starting from this statement, we investigate an optical method based on microcavities to perform single-cell characterization on bacteria. The analysis of the cell Brownian motion video in the trap combine with the optical structure transmission signal allow a fine characterization of the cell behavior. Moreover, by analysis the fluctuations of the transmitted signal over time during bacteria trapping, we expect to observe variations between bacteria phenotypes.

A 1D microcavity resonator is made on SOI substrate (250 nm / 2 μ m / 700 μ m) by combining electron beam lithography and reactive-ion etching. The two mirrors are designed for mode adaptation within the cavity³⁸ (Figure 2(a)) in order to get a quality factor of 4000. In this study we used the same optical structure for all the experiments. By injecting the resonance wavelength in this structure (laser at $\lambda_{\text{res}} = 1.5 \mu\text{m}$, injected power $P_{\text{inj}} = 10 \text{ mW}$, and linewidth $Lw = 10^{-3} \text{ pm}$), an intense and confined electromagnetic field is built in the cavity. The evanescent part of this field above the cavity creates optical gradient forces able to trap and manipulate colloidal objects.

Above the optical structures, a PDMS fluidic system leads swimming objects to the evanescent field. It consists of a

100 μm thick PDMS film with molded μ -fluidic channel in which we place a drop of water/bacteria solution. Then it is covered by a 150 μm thick layer of glass. This optofluidic chip is shown in Figure 2(b). A polarization-maintaining fiber enables the optical coupling of the light from a tunable laser to the microcavity. Two microscope objectives are placed on-top/in-line with the chip and connected to detection devices, which allows spatial (images) and temporal (signal) characterization of the trapping. Images of the cavity are created by a microscope connected to a CCD camera, with a time resolution of 1/24s. To track bacterium trajectories we detect their position on every recorded image for a relevant duration (few seconds), using tracking macro of Fiji (image processing package of ImageJ).³⁹ This analysis is shown in Figure 2(b-SPATIAL).

The signal transmitted by the optical structure is detected by a photodiode, increased by a trans-impedance amplifier and displayed on an oscilloscope. The oscillations observed Figure 2(b-TEMPORAL) are due to the shift of the resonance wavelength of the optical structure in presence of bacterium above the cavity. Indeed, the resonance wavelength depends not only on the optical index of the cavity, but also on the index of the environment. When the bacterium interacts with the electromagnetic field in the cavity, the environmental index ranges from $n_{\text{water}} = 1.33$ (bacterium outside the cavity, only water above it) to $1.368 < n_{\text{bacterium}} < 1.413$ (bacterium exactly above the cavity).⁴⁰ This phenomenon is shown in Figure 1: it induces a less than 1 nm shift of the resonance wavelength from the grey to the blue curve. As we keep injecting the same wavelength during all trapping experiments (resonance at n_{water}), the signal will oscillate between the two intensity values I_{MIN} and I_{MAX} while the bacterium oscillate between two positions: trapped above and untrapped around the cavity. By this method, we are able to record the position of the trapped object and the fluctuation of the transmitted signal simultaneously.

The objects under study are the following bacteria, shown Figure 2(c): *Staphylococcus epidermidis* diplococcus, *Escherichia coli* and *Bacillus subtilis* aggregate. They were selected for their size, close to the previously studied polystyrene beads^{30,41,42} and the differences in morphology and Gram-type between them (one Gram+ bacillus, one Gram- bacillus and one Gram+ coccus). An *E. coli* cell and a diplococcus of *S. epidermidis* have equivalent size and shape: oval shape and size of 1 μm x 2 μm , compared to a more elongated shape of 1 μm x 7 μm for *B. subtilis* (aggregate). *E. coli* and *B. subtilis* have flagella, a molecular motor they use to propel themselves through an aqueous medium. Bacterial strains (*E. coli* ATCC 11775, *S. epidermidis* ATCC 14990, *B. subtilis* ATCC 11774) were obtained from KwikStik lyophilized reference strains (Microbiologics, St. Cloud, MN).

Starting from a culture on agar media (COS or TSA, bioMérieux) after 24h of incubation (37°C), a 0.5 McF suspension (Densimat, bioMérieux) was prepared in API Suspension medium (bioMérieux).

We present now the two analysis methods of bacteria trapping, leading to their characterization.

	Bacterium	Confinement Factor $\frac{\mathcal{L}_B \times \ell_B}{\mathcal{L}_T \times \ell_T}$	Transversal confinement ratio ($\mathcal{L}_B / \mathcal{L}_T$)	Lateral confinement ratio (ℓ_B / ℓ_T)
(1)	<i>S. epidermidis</i>	57	9.5	6
(2)	<i>E. coli</i>	30.6	15.5	2
(3)	<i>B. subtilis</i>	3.2	2.8	1.1

Table 1. Numerical analysis of spatial behavior of trapped and non-trapped bacteria. The transversal direction is parallel to the x axis and the lateral one is parallel to the y axis.

The Brownian motion of each type of studied bacterium and the effect of trapping on its trajectory is shown in Figure 3. We characterize the effect of the trap by a parameter that we call ‘‘confinement factor’’ which consists in the ratio of the rectangular area covered by bacteria position ($\mathcal{L} \times \ell$) without and with trapping. The trapping is demonstrated by the reduction of the displacement area (confinement factor between 3.2 and 57) and its major effect is a transversal confinement (area reduced by a factor ranging from 2.8 to 15.5, in comparison to 1.1 to 6 for lateral confinement).

In this microcavity, previous studies have evidenced that the evanescent field is spread over two maxima of electromagnetic field.⁴² We observe in Figure 3(b) that *S. epidermidis* and *E. coli* interact with one maximum of the field at a time (*S. epidermidis* stays on one while *E. coli* keep switching from one maxima to another). This is allowed by their reduced size compared to the size order of the field maxima of the cavity. The flagella (*E. coli* and *B. subtilis*) seem to induce a larger displacement in the trap and the ability to interact with several maxima of the field. This behavior is consistent with the increased motility of the flagellated bacteria and with the shape of the electromagnetic field in the microcavity.

In Table 1, more details about confinement are shown. It highlights a stronger effect of the trap on *S. epidermidis* and *E. coli*, than on *B. subtilis* (confinement factor are 57, 30.6 and 3.2 respectively). This table also outlines a more effective confinement along transversal axis than along lateral one. For *S. epidermidis* lateral and transversal confinement are of the same order of magnitude, which can be linked to the spherical shape of the bacterium. On the other hand, for *E. coli* and *B. subtilis* the transversal confinement prevails. It is consistent with the behavior of the bacteria in the trap: they align on the structure, with their longer dimension parallel to the cavity. From this we can infer that the lateral and transversal contributions provide information on the shape of the object and its symmetry. The size and shape of *B. subtilis* induce only

transversal confinement (lateral confinement of 1.1), which explains the low effectiveness of the trap on that bacterium (confinement factor of 3.2) in accordance with experimental results (trapping duration under 10 minutes against more than 1 hour for *S. epidermidis* and *E. coli*). Finally, each type of the three investigated bacteria species owns a very distinct spatial signature, which is linked to their size, shape (symmetry) and flagella.

In parallel, monitoring the optical signal transmitted by the cavity (Figure 4) enables an accurate detection of the trapping effectiveness: high frequency oscillations and intensity drop of about 20% (Figure 4(a)).

On Figure 4(b) the temporal variation of the optical signal transmitted by the microcavity during the trapping of the three kinds of bacteria is shown. It is compared to the signal detected in the same conditions without trapping. Because the amplitude is the most relevant criteria to discriminate these curves, we calculate the histogram function of the transmitted signal (Figure 4(c)). Its characteristic elements are the full width at half maximum (FWHM) and the probability to have each intensity value. *E. coli* is characterized by a FWHM at least twice larger than the two other bacteria. It can be linked to its large lateral displacement in the trap (Figure 3(b, blue curve)). On the contrary, *S. epidermidis* trapping signal has a very low FWHM which highlights the unique position of equilibrium of the bacterium (high probability for one intensity value), matching with one field maximum. Finally, the optical signal transmitted by the adapted SOI microcavity is self-consistent for bacteria discrimination in this example. We highlighted here that amplitude is the most relevant criteria to differentiate the optical signal transmitted by the microcavity, in which the substantial part of the information is located below 15 Hz. It provides information on the maximum and minimum intensity values of the signal shown in Figure 1 and Figure 4. On Figure 5, these I_{max} and I_{min} values are represented on the resonance peak of the microcavity, at the injected wavelength.

If we consider that the quality factor remains constant during the trapping (low change of optical index when the microcavity environment is water or bacteria), it allows to calculate the redshift of the resonance wavelength between the two extreme positions of the bacteria in the trap (from the trap center (I_{min}) to its edge (I_{max})). This redshift is characteristic of bacterium phenotype, as represented Figure 5(b₁): its value ranges from 0.20 nm for *S. epidermidis* to 0.30 nm for *E. coli*.

Each colored peak represents the resonant mode available according to the type of trapped bacterium. By projecting these curves on the resonance peak of the optical trapping structure we are able to assign to every type of bacterium a specific area of the peak (Figure 5(b₂)) and discriminate them.

Finally, the two methods of characterization (spatial and temporal) are consistent with each other: the displacement of *E. coli* on the two evanescent field maxima can be linked to the large amplitude of its trapping signal and the higher value of the FWHM of its histogram function. *S. epidermidis* oscillates close to its balance position (one field maximum), matching to one transmitted intensity value of highest probability.

In this paper we show that planktonic single cell bacterium can be trapped by the evanescent field of a properly tuned microcavity. Both spatial (Brownian and partially-trapped motions) and temporal (optical transmission measurement) methods are relevant to study finely the bacterium behavior within the trap.

The spatial signature demonstrates that morphological properties of bacteria (size, shape and presence of flagella) can be identified thanks to the so-called confinement factor parameter.

The temporal signature through the analysis of the fluctuations of the optical transmitted signal over time during bacteria trapping allows us to identify bacteria phenotypes.

Compared to the spatial characterization, the temporal method requires only cavity transmission measurements. Thus, this promising label-free technic that can be performed in a much more compact way, with a smaller footprint, in a device scheme might be suitable for a reduce cost fast analysis device.

In the future we plan to extend this study to a larger number of bacteria, with different shapes, sizes and overall properties.

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Bibliography

- ¹ A. Ashkin and J.M. Dziedzic, Science **235**, 1517 (1987)
- ² J. Enger, M. Goksör, K. Ramser, P. Hagberg, and D.

Hanstorp, *Lab Chip* **4**, 196 (2004).

³ K. König, H. Liang, M.W. Berns, and B.J. Tromberg, *Opt. Lett.* **21**, 1090 (1996).

⁴ Y. Liu, G.J. Sonek, M.W. Berns, and B.J. Tromberg, *Biophys. J.* **71**, 2158 (1996).

⁵ K.C. Neuman, E.H. Chadd, G.F. Liou, K. Bergman, and S.M. Block, *Biophys. J.* **77**, 2856 (1999).

⁶ M. Ericsson, D. HANSTORP, P. HAGBERG, and J. ENGER, *J. Bacteriol.* **182**, 5551 (2000).

⁷ M.B. Rasmussen, L.B. Oddershede, and H. Siegumfeldt, *Appl. Environ. Microbiol.* **74**, 2441 (2008).

⁸ S. Mandal, J.M. Goddard, and D. Erickson, *Lab Chip* **9**, 2924 (2009).

⁹ S. Lin and K.B. Crozier, *ACS Nano* **7**, 1725 (2013).

¹⁰ M.R. Lee and P.M. Fauchet, *Opt. Express* **15**, 4530 (2007).

¹¹ S. Pal, P.M. Fauchet, and B.L. Miller, *Anal. Chem.* **84**, 8900 (2012).

¹² F. Vollmer, D. Braun, A. Libchaber, M. Khoshshima, I. Teraoka, and S. Arnold, *Appl. Phys. Lett.* **80**, 4057 (2002).

¹³ P. Kang, P. Schein, X. Serey, D. O'Dell, and D. Erickson, *Sci. Rep.* **5**, 12087 (2015).

¹⁴ F. Vollmer, S. Arnold, and D. Keng, *Proc. Natl. Acad. Sci. U. S. A.* **105**, 20701 (2008).

¹⁵ T. van Leest and J. Caro, *Lab Chip* **13**, 4358 (2013).

¹⁶ S. Chan, S.R. Horner, P.M. Fauchet, and B.L. Miller, *J. Am. Chem. Soc.* **123**, 11797 (2001).

¹⁷ P.B. Deotare, M.W. McCutcheon, I.W. Frank, M. Khan, and M. Lončar, *Appl. Phys. Lett.* **94**, 1 (2009).

¹⁸ M.W. McCutcheon and M. Loncar, *Opt. Express* **16**, 19136 (2008).

¹⁹ M. Notomi, E. Kuramochi, and H. Taniyama, *Conf. Proc. - Lasers Electro-Optics Soc. Annu. Meet.* **16**, 689 (2008).

²⁰ Y. Akahane, T. Asano, B. Song, and S. Noda, *Nature* **425**, 944 (2003).

²¹ Z. Zhang and M. Qiu, *Opt. Express* **12**, 3988 (2004).

²² J.B. Jager, V. Calvo, E. Delamadeleine, E. Hadji, P. Noé, T. Ricart, D. Bucci, and A. Morand, *Appl. Phys. Lett.* **99**, 2014 (2011).

²³ T.J. Kippenberg, S.M. Spillane, and K.J. Vahala, *Appl. Phys. Lett.* **85**, 6113 (2004).

²⁴ D.K. Armani, T.J. Kippenberg, S.M. Spillane and K.J.Vahala, **421**, 925 (2003).

²⁵ T. Baehr-Jones, M. Hochberg, C. Walker, and A. Scherer, *Appl. Phys. Lett.* **85**, 3346 (2004).

²⁶ I. Grudin, A. Savchenkov, A. Matsko, D. Strelakov, V. Ilchenko, and L. Maleki, *IQEC, Int. Quantum Electron. Conf. Proc.* **2005**, 380 (2005).

²⁷ M.L. Gorodetsky and V.S. Ilchenko, *J. Opt. Soc. Am. B* **16**, 147 (1999).

²⁸ L. Collot, V. Lefèvre-Seguin, M. Brune, J.M. Raimond, and S. Haroche, *Europhys. Lett.* **23**, 327 (2007).

²⁹ A. Rahmani and P.C. Chaumet, *Opt. Express* **14**, 6353 (2006).

³⁰ C. Pin, B. Cluzel, C. Renault, D. Peyrade, E. Picard, E.

Hadji, and F. De Fornel, *Appl. Phys. Lett.* **105**, 2012 (2014).

³¹ C. Renault, B. Cluzel, J. Dellinger, L. Lalouat, E. Picard, D. Peyrade, E. Hadji, and F. de Fornel, *Sci. Rep.* **3**, 2290 (2013).

³² S. Mandal, X. Serey, and D. Erickson, *Nano Lett.* **10**, 99 (2010).

³³ N. Deschamps, U.P. Dharanipathy, Z. Diao, M. Tonin, and R. Houdre, *Lab Chip* **13**, 3268 (2013).

³⁴ S. Lin, E. Schonbrun, and K. Crozier, *Nano Lett.* **10**, 2408 (2010).

³⁵ H. Cai and A.W. Poon, *Opt. Lett.* **35**, 2855 (2010).

³⁶ M.S. Luchansky and R.C. Bailey, *Anal. Chem.* **84**, 793 (2012).

³⁷ F. Vollmer, L. Yang, and S. Fainman, *Nanophotonics* **1**, 267 (2012).

³⁸ P. Velha, E. Picard, T. Charvolin, E. Hadji, J.C. Rodier, P. Lalanne, and D. Peyrade, *Opt. Express* **15**, 16090 (2007).

³⁹ J. Schindelin, I. Arganda-Carreras, E. Frise, V. Kaynig, M. Longair, T. Pietzsch, S. Preibisch, C. Rueden, S. Saalfeld, B. Schmid, J.-Y. Tinevez, D.J. White, V. Hartenstein, K. Eliceiri, P. Tomancak, and A. Cardona, *Nat. Methods* **9**, 676 (2012).

⁴⁰ J.B. Bateman, J. Wagman, and E.L. Carstensen, *Kolloid-Zeitschrift Zeitschrift F??r Polym.* **208**, 44 (1966).

⁴¹ C. Renault, J. Dellinger, B. Cluzel, T. Honegger, D. Peyrade, E. Picard, F. De Fornel, and E. Hadji, *Appl. Phys. Lett.* **100**, 1 (2012).

⁴² C. Pin, B. Cluzel, C. Renault, E. Picard, D. Peyrade, E. Hadji, and F. De Fornel, *ACS Photonics* **2**, 1410 (2015).

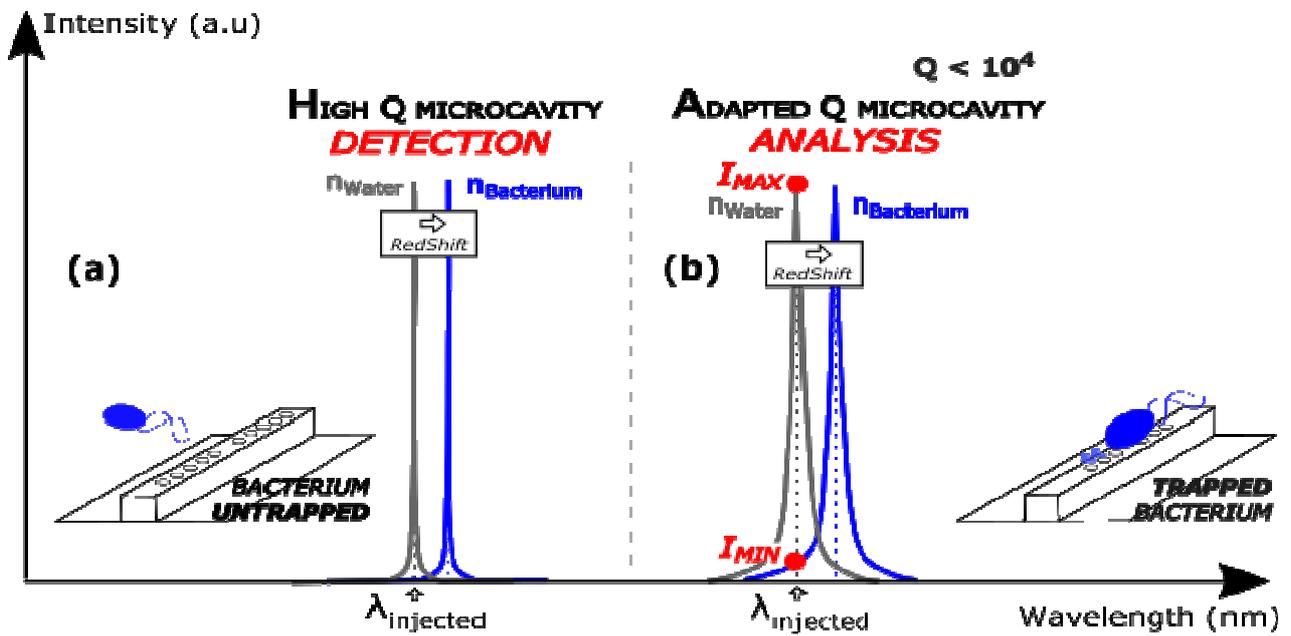


FIG. 1. Redshift of the microcavity resonance wavelength in presence of a bacterium above it. (a) High quality factor structure, ideal for detection of the target object, no trapping of bacteria. (b) Intermediate quality factor structure, ideal for bacteria trapping and analysis.

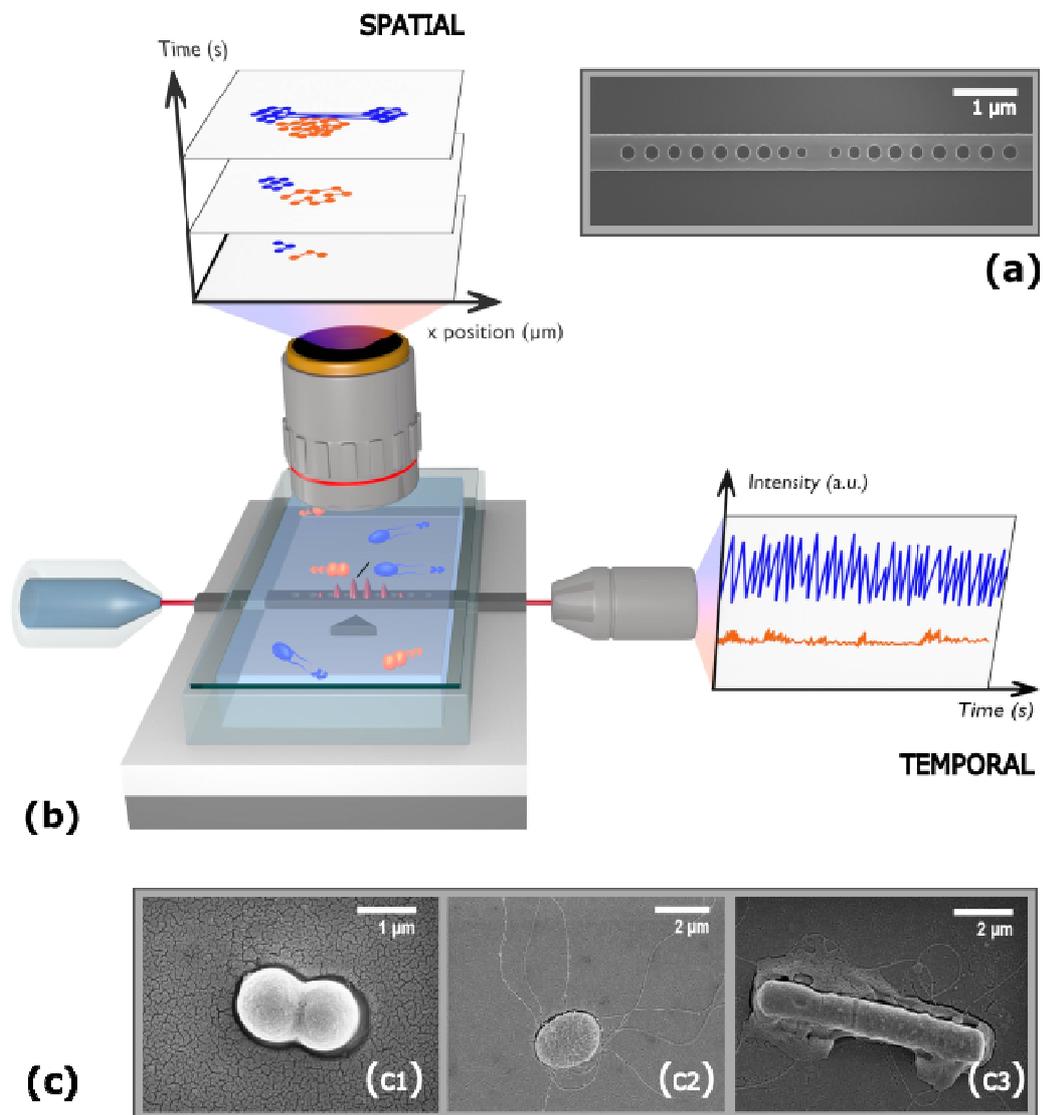


FIG. 2. (a) SEM image of the single microcavity used for all the experiments. (b) Trapping Set-up: fluidic system on a 1D PhC, light injected in the optical structure with a polarized fiber and recorded on structure output (TEMPORAL). Second trapping analysis way by microscopy column above the cavity (SPATIAL). The measurements related to *E. coli* are shown in blue and those associated with *S. epidermidis* are shown in orange, (c) SEM images of the under study bacteria: (1) *diplococcus* of *Staphylococcus epidermidis*, (2) *Escherichia coli* and (3) aggregate of *Bacillus subtilis*.

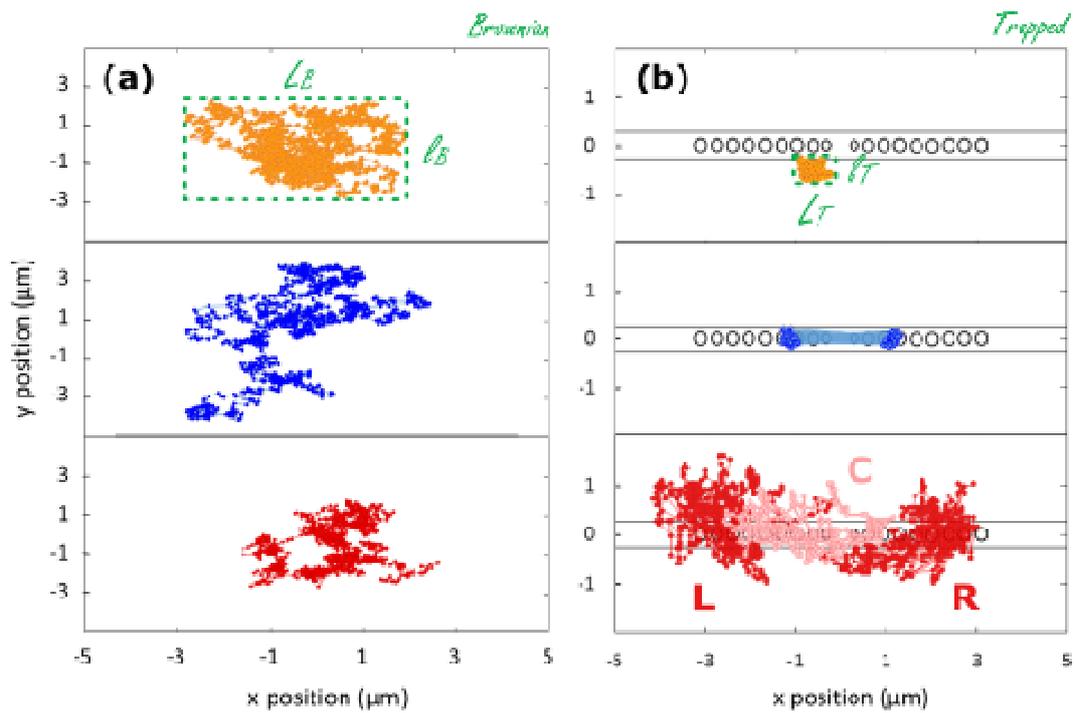


FIG. 3. (a) Brownian motion of a diplococcus of *S. epidermidis* (orange curve), a *E. coli* cell (blue curve) and a *B. subtilis* aggregate (red curve) during 50 seconds, tracking of bacteria center. (b) Trapped trajectories of *S. epidermidis* diplococcus (orange curve), *E. coli* (blue curve) and *B. subtilis* (red curve) during 50 seconds. Tracking of bacteria center for *S. epidermidis* and *E. coli*, tracking at left edge (L), center (C) and right edge (R) for *B. subtilis*.

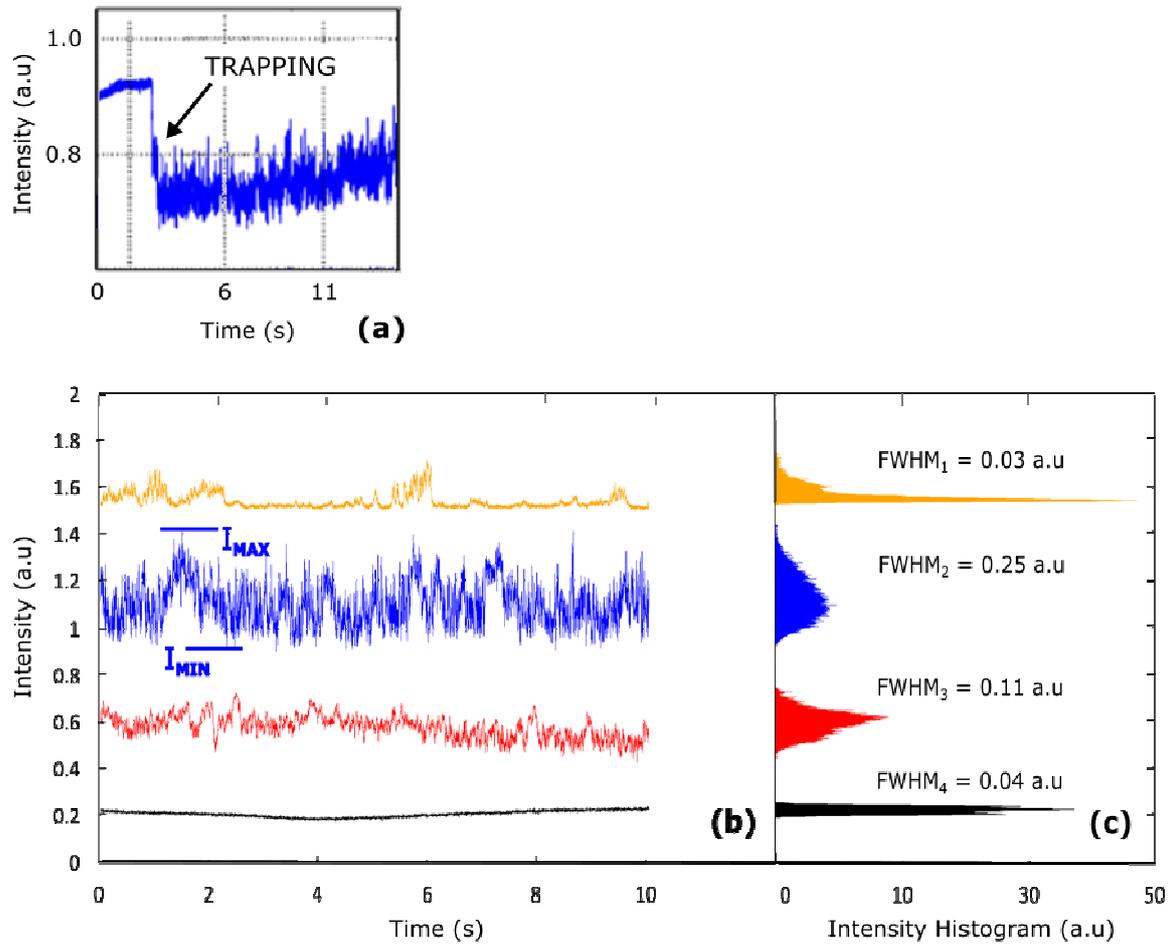


FIG. 4. (a) 20% drop in the optical signal transmitted by the microcavity, start of bacteria trapping. (b) Intensity transmitted by the microcavity during 10 seconds of trapping for the bacteria *S. epidermidis diplococcus* (orange), *E. coli* (blue), *B. subtilis aggregate* (red) and with no bacteria trapped (in bacteria and water medium) (black). (c) Histogram function of the transmitted optical signal.

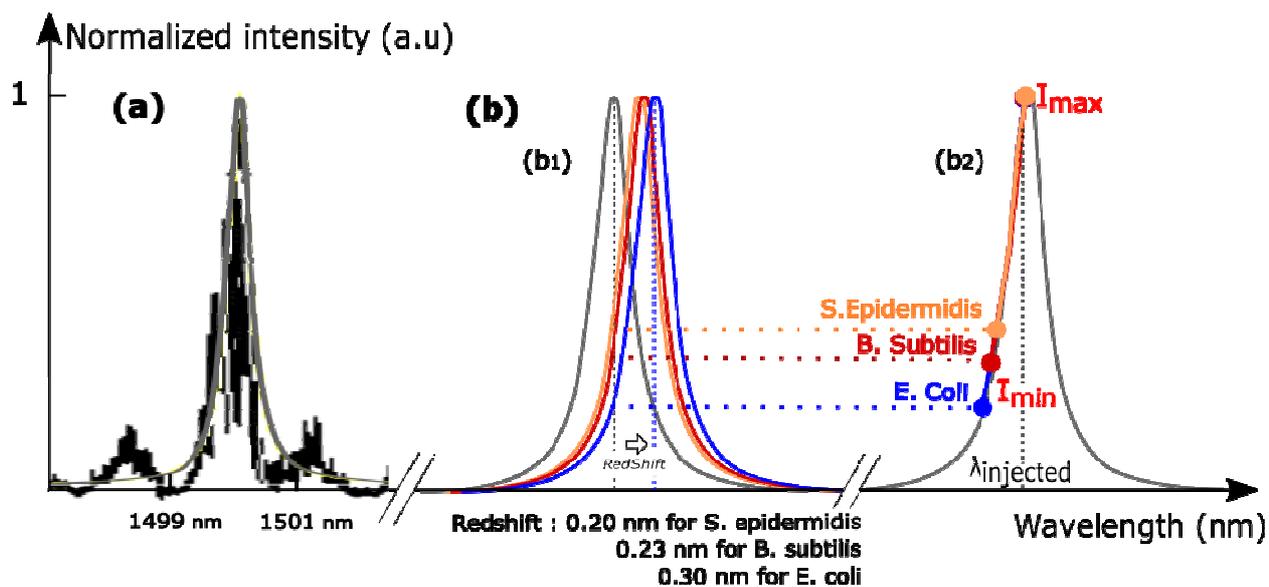


FIG. 5. (a) Experimental measurements, resonance peak of the optical structure used to trap bacteria (in black) and lorentzian fit of this curve (in grey), using Fityk (curve fitting and peak fitting software). (b) (1) Redshift of the microcavity resonance wavelength in presence of a diplococcus of *S. epidermidis* (orange curve), an aggregate of *B. subtilis* (red curve) and an *E. coli* cell (blue curve) above the optical trap. (2) Representation of the redshift of the resonance curve, delineated areas for each type of trapped bacteria. These curves were obtained by a lorentzian fit to experimental curves, at the right scale.